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Aspects of *Helicobacter pylori* transmission

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SUMMARY

The bacterium *Helicobacter pylori* infects the gastric mucosa of about half of the world's population. The infection causes gastritis and contributes to the development of peptic ulcer disease and gastric cancer. *H. pylori* infection is associated with low socioeconomic status, is typically acquired in early childhood and once established can persist throughout life unless treated. Person-to-person transmission appears to predominate and the family stands out as the primary framework for transmission.

In this thesis, an initial cross-sectional study aimed to disentangle the independent contributions of *H. pylori* infections in family members to the risk for the infection in 11- to 13-year old index children from Stockholm schools. *H. pylori* infections in mothers and in siblings, but not in fathers, were notable risk factors for infection in the index children. Furthermore, birth of the index child in a country with high *H. pylori* prevalence was an independent risk factor for infection. In addition to the initial standard analysis, a weighted logistic regression method was applied to accommodate additional non-randomly sampled cases. This exemplified how appropriate analysis of epidemiological data from complex sampling schemes can improve precision and maintain validity, while enabling a more complete investigation of risk factors already identified.

A subset of the infected family members underwent gastroscopy and contributed gastric biopsies from which *H. pylori* was isolated and typed by molecular methods. The same bacterial strains were frequently detected among siblings and between mothers and offspring. No strain concordance was detected between fathers and offspring, but parents sometimes harbored the same strains. The bacterial isolates were also examined with regard to the presence or absence of the *cag* pathogenicity island (PAI), a bacterial virulence factor. In a comparison with serological data, serology was supported as a suitable method to determine *cag* PAI status of *H. pylori* infections in clinical and epidemiological studies. Moreover, clonal and non-clonal bacterial isolates from members of a family were analyzed in more detail, which included microarray-based genome comparisons. Non-clonal *H. pylori* isolates exhibited extensive genetic variability, where certain characteristics could be discerned. However, transmission and host adaptation did not appear to be associated with substantial sequence diversity in the bacterial genome.

The present data support a predominantly mother-child and sib-sib transmission of *H. pylori*, consistent with an important role of intimate contact in the transmission. Furthermore, methodological and microbiological aspects that could aid future research are described. In summary, the findings of this thesis and the discussions thereof shed some light on the characteristics and mechanisms of transmission and persistence of *H. pylori* infection.

SAMMANFATTNING

Bakterien *Helicobacter pylori* infekterar magslemhinnan hos omkring hälften av världens befolkning. Infektionen orsakar gastrit och bidrar till utvecklandet av magsår samt magcancer. *H. pylori*-infektion är associerad med låg socioekonomisk status, inleds vanligen i tidig barndom och när den etablerats kan den utan behandling förbli livslång. Smittspridning tycks huvudsakligen ske från person till person och familjen synes utgöra den miljö där transmission framförallt äger rum.

I denna avhandling syftade en inledande tvärsnittsstudie till att särskilja oberoende effekter av *H. pylori*-infektion hos familjemedlemmar på infektionsrisken hos 11 till 13 år gamla indexbarn från skolor i Stockholmsområdet. *H. pylori*-infektion hos mödrar och syskon, men inte hos fäder, var betydande riskfaktorer för infektion hos indexbarnen. Dessutom var det en oberoende riskfaktor om indexbarnet var fött i ett land med hög prevalens av *H. pylori*. Utöver den inledande standardanalysen tillämpades en viktad logistisk regressionsmetod för att kunna inkludera ytterligare icke slumpmässigt utvalda fall. Detta exemplifierade hur en passande analys av epidemiologiska data från komplexa urvalsprocesser kan förbättra precision, bibehålla validitet samt möjliggöra en mer komplett utredning av redan identifierade riskfaktorer.

En del av de infekterade familjemedlemmarna genomgick gastroskopi och bidrog med biopsier från magslemhinnan, från vilka *H. pylori* isolerades och typbestämdes med molekylära metoder. Samma bakteriestammar identifierades ofta hos syskon samt mellan mödrar och barn. Ingen konkordans av stammar kunde detekteras mellan fäder och barn, men föräldrar var i vissa fall infekterade med samma stammar. De bakteriella isolaten undersöktes också med avseende på om de innehöll *cag* patogenicitetsön (PAI), som är en bakteriell virulensfaktor. I en jämförelse med serologiska data fann serologin stöd som en passande metod för att bestämma *cag* PAI-status hos *H. pylori*-infektioner i kliniska och epidemiologiska studier. Klonala och icke klonala bakteriella isolat från medlemmar i en familj studerades vidare i mer detalj, vilket innefattade microarray-baserade genomjämförelser. Icke klonala *H. pylori*-isolat uppvisade omfattande genetisk variabilitet, där vissa karakteristika kunde urskiljas. Däremot tedde sig transmission och värdanpassning inte vara associerade med betydande sekvensvariation i det bakteriella genomet.

Dessa data ger stöd för att *H. pylori* företrädesvis smittar från mödrar till barn samt mellan syskon, vilket är förenligt med att intim kontakt spelar en viktig roll i transmissionen. Dessutom beskrivs metodologiska och mikrobiologiska aspekter, vilka kan vara av relevans för framtida forskning. Sammanfattningsvis ger fynden i denna avhandling och diskussionen därav viss vägledning vad det gäller karakteristika och mekanismer för transmission och kronicitet av *H. pylori*-infektion.

LIST OF PUBLICATIONS

The thesis is based on the following papers, which will be referred to by their Roman numerals.

- Kivi M., Johansson A.L.V., Reilly M., Tindberg Y. *Helicobacter pylori* status in family members as risk factors for infection in children. *Epidemiology and Infection*. 2005(133): 645–652.
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- II. Kivi M., Johansson A.L.V., Salim A., Tindberg Y., Reilly M.
 Accommodation of additional non-randomly sampled cases in a study of *Helicobacter pylori* infection in families.
 Statistics in Medicine. In press. © John Wiley & Sons
- III. Kivi M., Tindberg Y., Sörberg M., Casswall T.H., Befrits R., Hellström P.M., Bengtsson C., Engstrand L., Granström M. Concordance of *Helicobacter pylori* strains within families. *Journal of Clinical Microbiology*. 2003(41): 5604–5608.
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- IV. Kivi M., Tindberg Y., Bengtsson C., Engstrand L., Granström M. Assessment of the *cag* pathogenicity island status of *Helicobacter pylori* infections with serology and PCR. *Clinical Microbiology and Infection*. 2005 (11): 66–68.
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- V. Kivi M., Hjalmarsson S., Kupershmidt I., Lundin A., Tindberg Y., Granström M., Engstrand L. *Helicobacter pylori* genome variability in a framework of familial transmission. *In manuscript*.

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LIST OF ABBREVIATIONS

CI	Confidence interval		
DI	Discriminatory index		
ELISA	Enzyme-linked immunosorbent assay		
GERD	Gastroesophageal reflux disease		
IL	Interleukin		
LPS	Lipopolysaccharide		
MALT	Mucosa-associated lymphoid tissue		
MAR	Missing-at-random		
NSAID	Non-steroidal anti-inflammatory drug		
OMP	Outer membrane protein		
OR	Odds ratio		
PAI	Pathogenicity island		
RAPD	Random amplified polymorphic DNA		
RFLP	Restriction fragment length polymorphism		
SE	Standard error		
SES	Socioeconomic status		
TLR	Toll-like receptor		
UBT	Urea breath test		

1. INTRODUCTION

Signs of *Helicobacter pylori* infection such as gram-negative gastric bacilli, gastric urease and epidemics of hypochlorhydria have been described since the late nineteenth century (Marshall, 2001). These observations could be better explained after Warren and Marshall in the early 1980's managed to culture a bacterium that was to be designated *Campylobacter pyloridis* (Marshall and Warren, 1984). In 1989, the genus *Helicobacter* was created and the bacterium received the name *Helicobacter pylori* (Goodwin *et al.*, 1989).

Half of the world's population is estimated to be infected with *H. pylori*, which makes it one of the most common bacterial pathogens in humans (Torres *et al.*, 2000). The infection is associated with low socioeconomic status (SES), is typically acquired in early childhood and once established can persist throughout life unless treated (Torres *et al.*, 2000). The association between *H. pylori* and gastritis was recognized early (Marshall and Warren, 1984) and virtually all infected individuals have later been confirmed to have a usually asymptomatic gastritis. A crucial role of the infection in peptic ulcer disease has been firmly established, which has enabled a paradigm shift in the treatment of ulcer patients (NIH Consensus Conference, 1994). A link between *H. pylori* and gastric cancer has also been demonstrated (IARC, 1994) and has been corroborated in subsequent studies (Ekström *et al.*, 2001; Uemura *et al.*, 2001). However, only 10–20% of infected individuals manifest severe complications and this selectivity in disease progression is inadequately understood (Blaser and Atherton, 2004; Suerbaum and Michetti, 2002).

The high *H. pylori* prevalence in many parts of the world, in conjunction with the pertinent links to disease, render the understanding of this infection an important public health issue.

2. OCCURRENCE AND TRANSMISSION

There are substantial differences in *H. pylori* prevalence between populations, originating in different probabilities of acquisition and persistence. The disentanglement of effects on acquisition from those on persistence requires longitudinal studies and thus, the common cross-sectional studies in this field do not separate these effects. Furthermore, risk factors may in some instances reflect both exposure and susceptibility to the infection. Epidemiological data of *H. pylori* infection are sometimes inconclusive or even conflicting, which may be a result of real variations between settings or methodological imperfections. Nonetheless, a number of features of more general character can be discerned in the epidemiology of *H. pylori* infection.

2.1. Prevalence and incidence

There are considerable differences in *H. pylori* prevalence between high-income and low-income countries. The prevalence in child populations ranges from below 10% to over 80% in high-income and low-income countries, respectively (Torres *et al.*, 2000). The infection is also associated with low SES within countries (Graham *et al.*, 1991; Malaty *et al.*, 1992; Malaty *et al.*, 1996). In the United States, for instance, a significantly lower prevalence was found in Caucasians (26%) compared to Hispanics (65%) and Afro-Americans (66%) (Malaty *et al.*, 1992). This dissimilarity was interpreted to reflect the different socioeconomic backgrounds of the groups. In a follow-up study, it was found that the difference in prevalence between Afro-Americans and Caucasians resulted from different seroconversion rates, although the rate of seroreversion could also have played a role (Malaty *et al.*, 2002).

H. pylori infection is usually acquired before the age of five (Granström *et al.*, 1997; Malaty et al., 2002; Mitchell et al., 1992). Annual incidence rates over 20% have been reported in early childhood in low-income countries (Glynn et al., 2002; Goodman et al., 2005). This is consistent with the rapidly increasing prevalence seen in children under the age of five years in many parts of the world (Klein *et al.*, 1994; Torres et al., 2000). In high-income countries, the incidence in early childhood is typically between 1% and 10% (Granström et al., 1997; Kumagai et al., 1998; Malaty et al., 2002), but can also reach as high as 20% (Goodman et al., 2005). Furthermore, infections that apparently clear spontaneously have been reported particularly in childhood (Goodman et al., 2005; Granström et al., 1997; Klein et al., 1994; Malaty et al., 2002). As many as 80% of childhood infections were eliminated by 11 years of age in Sweden (Granström et al., 1997) and by two years of age in Mexico and the United States (Goodman et al., 2005). In adults in high-income countries, the seroconversion rates tend to be about 0.5-1% per annum with slightly higher rates of seroreversion (Kumagai et al., 1998; Parsonnet, 1995; Veldhuyzen van Zanten et al., 1994). In low-income countries, the annual incidence in adults can be higher (Parsonnet, 1995) and reported rates of reinfection after H. pylori eradication have approached 20% in some studies (Hildebrand *et al.*, 2001; Soto *et al.*, 2003; Wheeldon *et al.*, 2005).

H. pylori prevalence is generally found to increase with age, reaching 20–50% in adult populations in Europe and North America (Bergenzaun *et al.*, 1996; Malaty *et al.*, 1992; Veldhuyzen van Zanten *et al.*, 1994). This pattern has been interpreted to partly reflect a birth-cohort phenomenon caused by a higher incidence in the past due to poorer living conditions and sanitation (Banatvala *et al.*, 1993; Parsonnet, 1995). Indeed, mathematical modeling has suggested that the infection will eventually disappear in high-income countries even without intervention (Rupnow *et al.*, 2000). But today, and in the foreseeable future, the infection remains highly prevalent in low-income regions of the world as well as in considerable portions of the populations in high-income countries.

2.2. Risk factors for infection

An obvious necessary cause for acquisition of *H. pylori* infection is exposure to the bacterium. The probability of exposure depends on the characteristics of the infective source and contact, but factors of the recipient host and the bacterium may also influence the probabilities of acquisition and persistence (Table 1). The bacterium has to overcome numerous barriers to successfully establish an infection in a new individual:

- Exit from an infected individual
- Transient survival outside the gastric niche
- Introduction into a new host
- Colonization of the new gastric mucosa
- Maintenance of the colonization

A predominantly person-to-person transmission has been postulated. This notion is based on the clustering of the infection in families (Drumm *et al.*, 1990; Goodman and Correa, 2000; Rocha *et al.*, 2003; Rothenbacher *et al.*, 2002b) and in institutionalized individuals (Lambert *et al.*, 1995), while consistent and verified environmental reservoirs are absent (see "Environmental and behavioral factors" and "Transmission routes").

The family

The family stands out as the most important framework for transmission and a child's risk of being infected is associated with having infected family members (Goodman and Correa, 2000; Rocha *et al.*, 2003; Rothenbacher *et al.*, 2002b). Family size and residential crowding (persons per room or m^2) are frequently described as risk factors for *H. pylori* infection and may be regarded as proxies for the number of infected family members (Goodman *et al.*, 1996; Goodman and Correa, 2000; McCallion *et al.*, 1996; Mendall *et al.*, 1992; Tindberg *et al.*, 2001b; Webb *et al.*, 1994). Similarly, having familial connections to high-prevalence regions

Necessary cause: Exposure to the	References
Living in or originating from high-prevalence areas	(Rothenbacher <i>et al.</i> , 1998b; Tindberg <i>et al.</i> , 2001b; Torres <i>et al.</i> , 2000; Tsai <i>et al.</i> , 2005)
Large family size, infected family members	(Goodman <i>et al.</i> , 1996; Goodman and Correa, 2000; McCallion <i>et al.</i> , 1996; Mendall <i>et al.</i> , 1992; Rocha <i>et al.</i> , 2003; Rothenbacher <i>et al.</i> , 2002b; Tindberg <i>et al.</i> , 2001b; Webb <i>et al.</i> , 1994)
Infected contacts in the community: Daycare centers	(Dore <i>et al.</i> , 2002)
Environmental reservoirs: Contaminated water	(Brown <i>et al.</i> , 2002; Goodman <i>et al.</i> , 1996; Klein <i>et al.</i> , 1991; Nurgalieva <i>et al.</i> , 2002)
Behavior and other factors increasing the	(Brown <i>et al.</i> , 2002; Laporte <i>et al.</i> , 2004;
exposure: Intimate contact, gastroenteritis, poor sanitary practices	Luzza <i>et al.</i> , 2000; McCallion <i>et al.</i> , 1996; Nurgalieva <i>et al.</i> , 2002; Rocha <i>et al.</i> , 2003; Rothenbacher <i>et al.</i> , 2002b; Tindherg <i>et al.</i> , 2001b; Webb <i>et al.</i> , 1994)
Component cause: Host factors	References
Expression of receptors	(Aspholm-Hurtig <i>et al.</i> , 2004; Borén <i>et al.</i> , 1993; Rothenbacher <i>et al.</i> , 2004)
Host defenses: Gastric acid secretion,	(Björkholm <i>et al.</i> , 2004; Hartland <i>et al.</i> ,
immune responses	2004; Magnusson <i>et al.</i> , 2001; Mohammadi <i>et al.</i> , 1997)
Other factors affecting the gastric milieu: Young age, diet	(Granström <i>et al.</i> , 1997; Kuepper-Nybelen <i>et al.</i> , 2005; Malaty <i>et al.</i> , 2002; Mitchell <i>et al.</i> , 1992)
Component cause: Bacterial factors	References
Protected localization: Motility, adhesion, internalization	(Aspholm-Hurtig <i>et al.</i> , 2004; Björkholm <i>et al.</i> , 2000; Eaton <i>et al.</i> , 1992; Ilver <i>et al.</i> , 1998; Mahdavi <i>et al.</i> , 2002; Terry <i>et al.</i> ,
Withstanding host defenses: Urease activity, immune evasion	 2005) (Allen, 2001; Andersen-Nissen <i>et al.</i>, 2005; Bergman <i>et al.</i>, 2004; Boncristiano <i>et al.</i>, 2003; Bäckhed <i>et al.</i>, 2003; Eaton and Krakowka, 1994; Gebert <i>et al.</i>, 2003:
Adaptive evolution	Molinari <i>et al.</i> , 1998) (Aspholm-Hurtig <i>et al.</i> , 2004; Salaün <i>et al.</i> , 2005; Yamaoka <i>et al.</i> , 2002)

Table 1. Overview of possible determinants of *H. pylori* infection

is associated with infection in children living in low-prevalence areas (Rothenbacher *et al.*, 1998b; Tindberg *et al.*, 2001b) and this effect decreases in successive generations (Tsai *et al.*, 2005). The living conditions during childhood can be predictive of infection in adulthood (Mendall *et al.*, 1992; Webb *et al.*, 1994; Woodward *et al.*, 2000), being in accordance with *H. pylori* acquisition in childhood

from household members. Furthermore, the possibility of child-child transmission outside the family was not supported in a Swedish study, where the *H. pylori* prevalence in classmates was not a risk factor for infection (Tindberg *et al.*, 2001b). However, daycare attendance was associated with *H. pylori* infection in urban Sardinian children (Dore *et al.*, 2002).

Having an infected mother has been found to be a more prominent risk factor for childhood infection than having an infected father, supporting primarily mother-child transmission (Rocha *et al.*, 2003; Rothenbacher *et al.*, 2002b; Tindberg *et al.*, 2001b). Transmission among siblings has also been indicated by clustering of the infection in sibships (Goodman and Correa, 2000; Rocha *et al.*, 2003). Data from a high-prevalence area in Colombia suggested that having older infected siblings (Goodman and Correa, 2000). A narrower age gap to the next older infected sibling also seemed to increase the risk for infection. The importance of both infected mothers and siblings has thereafter been corroborated in a high-prevalence community in Brazil (Rocha *et al.*, 2003). The suggested central roles of infected mothers and siblings, and the lesser role played by infected fathers, probably reflect how intimate contact potentiates the effect of having infected family members.

Even though the infection is usually initiated in early childhood, some epidemiological data point to the possibility of acquisition in adulthood from infected family members. Having an infected spouse has been described as a risk factor for infection (Brenner *et al.*, 1999). This study controlled for the country of origin, which could otherwise have explained the association (Perez-Perez *et al.*, 1991). Furthermore, having more children has been identified as a risk factor for infection in adults (Mendall *et al.*, 1992), possibly indicating that children may serve as mediators of transmission within families. Some additional, albeit weak, evidence in this direction may be that having an infected spouse was not a risk factor for infection in childless couples (Perez-Perez *et al.*, 1991).

Environmental and behavioral factors

Reasons for the association between *H. pylori* infection and low SES have been sought among environmental and behavioral factors. A shared environmental source of the infection could theoretically contribute to the observed intrafamilial clustering. Possibly contaminated water has been suggested as an infection source since using particular water sources, such as wells, has been correlated to the infection (Brown *et al.*, 2002; Goodman *et al.*, 1996; Klein *et al.*, 1991; Nurgalieva *et al.*, 2002). However, other studies have not found the water source to be associated with infection (Clemens *et al.*, 1996; Glynn *et al.*, 2002; Malaty *et al.*, 1996). More indirect environmental transmission has also been proposed following the identification of the consumption of raw vegetables as a risk factor for infection (Goodman *et al.*, 1996; Hopkins *et al.*, 1993). Furthermore, *H. pylori* has been proposed to possess zoonotic potential and suggested reservoirs include cats (Handt *et al.*, 1994), houseflies (Grübel *et al.*, 1997) and sheep (Dore *et al.*, 1999), but these theories are controversial.

Behavioral factors may influence the risk of *H. pylori* acquisition and persistence. Residence in a high-prevalence country can facilitate acquisition, as frequent close contact with infected individuals and poor sanitary practices may enhance bacterial exposure (Brown et al., 2002; Goodman et al., 1996; Hopkins et al., 1993; Nurgalieva et al., 2002). The importance of intimate contact for acquisition was mentioned above as a possible explanation for infected mothers and siblings, and not infected fathers, being primary determinants for the infection in children. Intimate contact has likewise been suggested to explain other observed risk factors, such as bed sharing (McCallion et al., 1996; Webb et al., 1994) and breastfeeding (Rothenbacher et al., 2002a). Breastfeeding has also been speculated to possibly provide protection against early infection by passive immunization (Blecker et al., 1994; Thomas et al., 1993). However, such protection, if any, should be of limited relevance after weaning, as supported by negative findings (Dore et al., 2002; Glynn et al., 2002; McCallion et al., 1996). Moreover, H. pylori infection has been negatively correlated with antibiotic consumption (Mitchell et al., 1992; Rothenbacher et al., 1998a). In another study, however, a similar negative association disappeared when the country of origin was taken into account, which could be explained by the higher antibiotic consumption in low-prevalence countries (Tindberg et al., 2001b). Some behavioral factors have been assessed as determinants for *H. pylori* infection specifically in adults. Examples include a possible negative association with alcohol intake (Kuepper-Nybelen et al., 2005) and perhaps a positive relationship with smoking (Woodward et al., 2000), but data are discrepant (Malaty et al., 1996; Mitchell et al., 1992; Woodward et al., 2000).

As illustrated above, data regarding environmental and behavioral factors as determinants for *H. pylori* infection are often inconclusive. The independent effects may be modest and confounding by other socioeconomic factors or household characteristics is a major concern. The importance of different risk factors may also differ between populations. Thus, the data on environmental and behavioral exposures are often complicated to interpret, but they may contain important information.

Host and bacterial factors

H. pylori strains differ in their ability to establish and maintain an infection in a given host, which can be attributed to host and bacterial factors and their compatibility (Aspholm-Hurtig *et al.*, 2004; Dubois *et al.*, 1999; Salaün *et al.*, 2005; Suto *et al.*, 2005; Yamaoka *et al.*, 2002). The transient infections in childhood may reflect instances where the bacterium is not optimally suited for the new host and adaptation is not feasible or rapid enough, leading to the host succeeding in clearing the infection (Goodman *et al.*, 2005; Granström *et al.*, 1997; Klein *et al.*, 1994; Malaty *et al.*, 2002).

Host genetics have been indicated to be involved in susceptibility to *H. pylori* infection, based on a higher concordance of infection in monozygotic (81%) than in dizygotic (63%) twin pairs (Malaty *et al.*, 1994). The specific genetic components of this suggested predisposition are unknown, but some host factors that may contribute

susceptibility to the infection have been proposed. Expression of blood group antigens that mediate bacterial adherence to the gastric mucosa has been suggested to be important for susceptibility to *H. pylori* infection (Borén *et al.*, 1993). Some research indicates that *H. pylori* strains have adapted their binding affinities in accordance with the blood group antigen expression of different human populations (Aspholm-Hurtig *et al.*, 2004). Furthermore, individuals that excrete receptors in body fluids, offering removable binding sites that can compete with the tissue-bound receptors, have been reported to have a lower risk of being infected (Rothenbacher *et al.*, 2004). However, some studies have shed doubt on the theory that blood group antigen-mediated adhesion contributes to susceptibility to infection (Clyne and Drumm, 1997; Umlauft *et al.*, 1996; Yamaoka *et al.*, 2002). These discrepancies may reflect multifaceted mechanisms of bacterial adhesion and a need for sophisticated measurements when assessing the binding potential.

The immune system may also be involved in determining predisposition to *H. pylori* infection. This notion is supported by studies that describe alleles within the human leukocyte antigen locus HLA-DQA1 to be correlated with the infection (Azuma *et al.*, 1995; Magnusson *et al.*, 2001). However, there are discordant data showing no association (Karhukorpi *et al.*, 1999). An interleukin (IL)-1 β receptor polymorphism of unknown functional consequence has also been correlated with the infection (Hartland *et al.*, 2004). Furthermore, the spread of the bacterium may be promoted by low gastric acid secretion, which may be especially relevant in young children and during infective gastroenteritis (Björkholm *et al.*, 2004; Cook, 1985). Some studies have found a slightly higher prevalence of *H. pylori* infection in males (Goodman *et al.*, 1996; Woodward *et al.*, 2000). The reasons for this tendency are unclear and other studies have not been able to confirm this correlation (Hopkins *et al.*, 1993; Malaty *et al.*, 2002; Mitchell *et al.*, 1992; Tindberg *et al.*, 2001b).

H. pylori has developed a repertoire of functions for survival in the harsh gastric niche, including acid tolerance, motility, adherence, immune evasion and mechanisms for adaptive evolution (see "The microorganism"). These features are all involved in the interplay between the host and the bacterium and may influence acquisition and persistence of infection, as is typically studied in animal models. Bacterial acid tolerance and motility were among the first factors found to play a role in colonization (Eaton et al., 1992; Eaton and Krakowka, 1994). Global mutagenesis approaches have verified these findings and have expanded the collection of putative essential genes (Kavermann et al., 2003; Salama et al., 2004). However, the relevance of these observations in human populations is largely unknown. In a Finnish population, strains with a bacterial virulence factor, the cag pathogenicity island (PAI), have been indicated to disappear more rapidly than strains without the cag PAI (Perez-Perez et al., 2002). This could speculatively be explained by reduced transmissibility or persistence of cag PAI+ strains in this population. However, cag PAI+ infections constitute the majority of H. pylori infections worldwide (Nilsson et al., 2003; Park et al., 2002; Parsonnet et al., 1997; Simán et al., 2005) and murine studies speak against a significant role of the cag PAI in colonization, at least after

the initial phase of the infection (Marchetti and Rappuoli, 2002; Yamaoka et al., 2002).

2.3. Molecular typing

Molecular typing has shown that unrelated individuals harbor distinct *H. pylori* strains (Akopyanz *et al.*, 1992; Taylor *et al.*, 1995). Nevertheless, *H. pylori* genetic variation has been correlated to different human populations and to human migrations such as European colonization and the slave trade (Achtman *et al.*, 1999; Falush *et al.*, 2003; Kersulyte *et al.*, 2000; Wirth *et al.*, 2004). Molecular typing can further exploit the bacterial genome diversity to study the spread of *H. pylori* within shorter time frames. The presence of genetically related bacteria in different individuals indicates person-to-person transmission or acquisition from a common source. Accordingly, molecular typing can corroborate and further characterize the transmission pathways suggested by epidemiological data based on infection status. In this respect, microbial typing techniques should always be evaluated for their ability to cluster related strains and discriminate between unrelated strains (Hunter and Gaston, 1988).

The terminology of bacterial samples and their relatedness is sometimes inconsistent, which justifies a clarification of the terminology that will be used here. An "isolate" is a collection of cells derived from a single cell (van Belkum *et al.*, 2001). A "strain" represents an isolate or a group of isolates displaying specific characteristics that set it apart from other isolates belonging to the same species. The term strain is commonly used interchangeably with "clone", that is, organisms descending from a common ancestor through a direct chain of replication.

H. pylori clonality can be discerned for isolates from different family members, occasionally in combination with clonal variants (Han et al., 2000; Raymond et al., 2004; van der Ende et al., 1996). These observations support intrafamilial transmission and bear analogy to an epidemic clonal population structure of microorganisms (van Belkum et al., 2001), although H. pylori is generally regarded as panmictic (Achtman et al., 1999; Suerbaum et al., 1998). It is difficult to obtain gastric biopsies from children and asymptomatic individuals and thus, familial molecular typing data are sparse and often based on small samples. Nevertheless, clustering of strains in sibships has been reported (Han et al., 2000; Miehlke et al., 1999; van der Ende et al., 1996) and it has been suggested that children are colonized more frequently with the mother's strain than the father's (Han et al., 2000). However, both father-offspring and mother-offspring strain concordance have been described (Bamford et al., 1993; Gibson et al., 1998; van der Ende et al., 1996). Some acquisition of the infection in adulthood is indicated by findings of shared strains among spouses (Bamford et al., 1993; Gibson et al., 1998; van der Ende et al., 1996) and reinfections with apparently new strains after H. pylori eradication (Hildebrand et al., 2001; Soto et al., 2003). Hence, the molecular typing data seem to support the transmission patterns outlined from other types of epidemiological studies.

H. pylori infection in an individual appears to be comprised of mainly one strain (Berg *et al.*, 1997; Raymond *et al.*, 2004; Taylor *et al.*, 1995). More than one strain can, however, be present and such mixed infections have been hypothesized to be more common in high-prevalence settings due to repeated acquisition opportunities (Berg *et al.*, 1997). Furthermore, the extensive genetic diversification of *H. pylori* renders each isolate derived from a single cell more or less unique and has lead to descriptions of the infection as being composed of a multitude of quasispecies (Blaser and Berg, 2001; Kuipers *et al.*, 2000). The presence of different strains and strain variants constitutes a methodological limitation in typing studies and has to be considered in the corresponding interpretations. A vast area of the gastric mucosa usually remains unsampled and some isolates may be overlooked. Moreover, quasispecies could perhaps mutate beyond the recognition of clonality by some molecular typing techniques.

2.4. Transmission routes

Possible *H. pylori* transmission routes are gastro-oral, fecal-oral or oral-oral, but firm evidence is scarce. *H. pylori* has been cultured from vomitus, diarrheal stools and saliva, demonstrating that the bacterium is potentially transmissible through these routes (Leung *et al.*, 1999; Parsonnet *et al.*, 1999; Thomas *et al.*, 1992). In contrast, *H. pylori* cultures of environmental water samples have only very rarely been successful (Lu *et al.*, 2002). Bacterial DNA can be detected in the environment by PCR (Hultén *et al.*, 1996), but the DNA may very well represent remnants of dead bacteria and does not constitute strong evidence for the possibility of environmental transmission.

Vomitus in particular appears to harbor viable bacteria and even air in the vicinity of a vomiting study subject can be *H. pylori* positive by culture (Parsonnet *et al.*, 1999). In line with this observation is a study which found a weak association between vomiting in siblings and childhood infection (Luzza et al., 2000). An elevated incidence of H. pylori infection after outbreaks of gastroenteritis has also been reported in a French institution (Laporte et al., 2004). Hepatitis A is spread through the fecal-oral route and correlations between antibodies against hepatitis A and H. pvlori could possibly indicate a common mode of transmission. However, a number of studies have not been able to confirm this hypothesis (Furuta et al., 1997; Hazell et al., 1994) and more conclusive evidence has to be sought elsewhere. Oral-oral transmission of H. pylori may be interpreted to be of limited importance, based on data showing that the prevalence of the infection was not higher in dentists compared to non-clinical colleagues (Matsuda et al., 2002), while it was higher in gastroenterologists compared to internists, nurses and population controls (Lin et al., 1994). Furthermore, episodes of diarrhea have been proposed to be more common in children during the months after H. pylori acquisition relative to uninfected and

persistently infected children, which may promote dissemination of the infection (Passaro *et al.*, 2001).

It is plausible that close contact within families facilitates exposure to bacteria through contaminated body excretions, being in agreement with familial transmission. Regurgitation, vomiting and diarrhea are common in childhood and children may boost familial *H. pylori* transmission when the bacterium is introduced into a child. Following this line of reasoning, it is appealing to envision a model in which societal development involves a decreasing frequency of gastrointestinal illnesses and improved sanitation, thereby contributing to the declining *H. pylori* prevalence in high-income parts of the world.

3. THE MICROORGANISM

"From our anthropocentric point of view, we say a person with the agent is infected. From the point of view of the infectious agent, however, humans are simply home and lunch, their ecologic niche." (Halloran, 1998)

3.1. General aspects of H. pylori microbiology and infection

H. pylori is the best known member of the *Helicobacter* genus, which includes dozens of species that primarily colonize the gastrointestinal tract of a variety of animals (Fox, 2002). *H. pylori* is a curved gram-negative bacillus with a bundle of unipolar flagella (Figure 1). Biochemical identification of *H. pylori* relies on the activities of the urease, catalase and oxidase enzymes. The bacterium is slow-growing and requires a rich medium and a microaerophilic atmosphere for *in vitro* culture. After starvation through prolonged culturing, a coccoid form can be found in the cultures and it has been debated whether this form represents dormant or degenerated, non-viable bacteria (Andersen *et al.*, 2000).



Figure 1. *H. pylori.* The curved bacillus with unipolar flagella is visualized by a scanning electron microscope (left) and depicted in a schematic drawing (right). The microscopic image was kindly provided by Christina Nilsson.

The human stomach is an inhospitable milieu and a fasting stomach is normally devoid of bacterial species other than *H. pylori* and some *Lactobacilli*. *H. pylori* is well adapted to its gastric niche and has developed a broad spectrum of functions that enable colonization. For example, bacterial urease hydrolyzes urea with the formation of carbon dioxide and ammonia, providing protection against the highly acidic gastric environment. The capability of *H. pylori* to maintain a chronic infection is of particular interest and can be facilitated by i) protected localization and adherence, ii) evasion and regulation of the immune response and iii) adaptation to changing conditions.

The infection can be patchy and is primarily localized to the distal parts of the stomach, but can spread proximally, especially in persons with low gastric acid secretion (Bayerdörffer *et al.*, 1989; Testerman *et al.*, 2001). The majority of the bacteria are regarded to be free-living in the gastric mucus layer, which can provide some protection against the harsh environment (Testerman *et al.*, 2001). Part of the bacterial population adheres to the gastric epithelial cells, which may benefit interactions with the host and maintenance of the colonization. A significant role of adherence is indicated by the array of products that contribute to this purpose in *H. pylori* (Edwards *et al.*, 2000; Ilver *et al.*, 1998; Mahdavi *et al.*, 2002; Testerman *et al.*, 2001; Yamaoka *et al.*, 2002). The bacterium is generally extracellular, but may invade host cells, although the significance of internalization is uncertain (Björkholm *et al.*, 2000; Testerman *et al.*, 2001).

Long-term persistence of the infection is likely to require evasion or modulation of the immune response. However, the inflammation may benefit the bacteria to some extent by disrupting the tissue integrity, thereby making nutrients available. Thus, bacterial interactions with the immune system may need to somewhat balance the risk of eradication against a more favorable environment (Blaser and Berg, 2001). The innate immune response towards *H. pylori* may be impaired by relatively inert interactions with the Toll-like receptors (TLR) (Andersen-Nissen *et al.*, 2005; Blaser and Atherton, 2004; Bäckhed *et al.*, 2003) and by resistance to phagocytic killing (Allen, 2001). The adaptive immune response is shifted towards a Th1 response, which is unusual for extracellular pathogens and may contribute to the inflammation (Blaser and Atherton, 2004; Mohammadi *et al.*, 1997). The bacterium can interact with cells of the adaptive immune system directly or through effector molecules to balance the Th1/Th2 responses (Bergman *et al.*, 2004), interfere with antigen presentation (Molinari *et al.*, 1998) and to inhibit activation or induce apoptosis of T lymphocytes (Boncristiano *et al.*, 2003; Gebert *et al.*, 2003; Wang *et al.*, 2001).

H. pylori is a genetically diverse bacterial species (Achtman *et al.*, 1999; Akopyanz *et al.*, 1992; Björkholm *et al.*, 2001b; Israel *et al.*, 2001; Salama *et al.*, 2000; Suerbaum *et al.*, 1998). Genetic diversification may facilitate immune evasion and adaptation to different gastric niches, new hosts or a changing gastric environment over the years. Furthermore, strain variants in the same stomach could constitute a reservoir of better adapted strains if the living requirements would change.

3.2. The genome

H. pylori is the first bacterial species for which two genomes, those of strains 26695 (Tomb *et al.*, 1997) and J99 (Alm *et al.*, 1999), were completely sequenced. A third sequenced genome, AG7:8, is currently in the final stages of annotation and analysis (H. Kling Bäckhed, personal communication). The *H. pylori* genome is small and compact and the 1.65 million base pairs accommodate about 1,500 genes. The limited metabolic potential and the low number of regulatory networks have been interpreted to reflect a restricted gastric niche of the bacterium (Doig *et al.*, 1999;

Tomb *et al.*, 1997). After a revision of the annotation, 77% of the genes have been assigned a functional category (Boneca *et al.*, 2003). Comparison of the two sequenced genomes revealed some larger genomic rearrangements, but the gene order and metabolic potential were relatively conserved (Alm *et al.*, 1999; Doig *et al.*, 1999). Strain-specific genes, 6-7% of the genes, were concentrated in two genomic regions that hence were designated "plasticity zones". These zones have thereafter also been discernible by microarray-based comparative genomics of other strains (Björkholm *et al.*, 2001a; Salama *et al.*, 2000). Some gene functional classes have been found to be especially variable, including genes related to DNA metabolism and the cell envelope (Boneca *et al.*, 2003; Salama *et al.*, 2000; Salaün *et al.*, 2005). Possible explanations for this variability are that DNA diversification and its regulation have important roles in *H. pylori* and that surface structures exposed to the host undergo antigenic variation. As described below, *H. pylori* has developed strategies to facilitate and regulate its extensive genetic diversification, probably to attain optimal instruments for adaptive evolution.

Generation of genome diversity

Recombination has been postulated to be a primary source of *H. pylori* genome diversity and panmixis (Achtman *et al.*, 1999; Falush *et al.*, 2001; Suerbaum *et al.*, 1998). One study estimated characteristics related to recombination by sequencing of DNA fragments from paired isolates (Falush *et al.*, 2001). The resulting estimates give a picture of frequent recombination of comparably small fragments (60 imports spanning 25,000 base pairs per genome per year, that is, 1.5% of the genome). However, the estimated rate of recombination may be inflated since only one isolate from each time point was considered and thus, the diversity could have been present initially. The genetic divergence may be slower in some instances, as discussed after the finding of limited differences in sequential isolates separated by nine years (Lundin *et al.*, 2005).

Horizontal genetic transfer is common in *H. pylori* and transformation, uptake of naked DNA from the environment, is facilitated by the widespread natural competence of *H. pylori* (Nedenskov-Sörensen *et al.*, 1990). Other means of horizontal genetic transfer are conjugation and phage-mediated transduction, of which the former DNase-resistant mechanism has been described in *H. pylori* (Kuipers *et al.*, 1998). The possibility of generating genome diversity by DNA transfer has been hypothesized to decrease in high-income countries due to the reduced circulation of different *H. pylori* strains (Blaser and Atherton, 2004). If this leads to less genetic diversity it may be a contributing factor to the declining *H. pylori* prevalence in high-income countries. Deletion of DNA segments also adds to the genetic diversity of *H. pylori*. In 15 unrelated isolates, 12–18% of the genes of 26695 and J99 were found to be dispensable in each isolate (Salama *et al.*, 2000), while 0.3–1.5% of the J99 genes were deemed to be absent in each of 13 J99 variants (Israel *et al.*, 2001).

Restriction-modification systems can control horizontal genetic transfer and a distinctive feature of the *H. pylori* genome is the presence of a high number of these

systems (Alm *et al.*, 1999; Lin *et al.*, 2001; Tomb *et al.*, 1997; Xu *et al.*, 2000). Functional restriction-modification systems consist of a methyltransferase that methylates a specific target DNA sequence and an endonuclease that cleaves the same sequence when it is unmethylated. Accordingly, invading DNA will be unmethylated and digested, while endogenous DNA will be methylated and protected. The *H. pylori* restriction-modification systems are variable and each strain contains its unique setup, which can sustain genome integrity and allow the existence of separate gene pools (Alm *et al.*, 1999; Lin *et al.*, 2001; Tomb *et al.*, 1997; Xu *et al.*, 2000). Furthermore, these systems may be conceived of as selfish DNA and may provide repetitive DNA and DNA breakage that could facilitate recombination or they may be involved in gene regulation by methylation of promotor sequences (Blaser and Berg, 2001).

H. pylori is prone to point mutation and a considerable proportion of strains have exceptionally high mutation rates, comparable to mutator strains in other species (Björkholm *et al.*, 2001b). A mutator phenotype indicates that the strain is deficient in DNA repair and *H. pylori* appears to lack components of the mismatch and SOS repair systems (Björkholm *et al.*, 2001b; Tomb *et al.*, 1997). Most point mutations do, however, occur at the third base of the codon, where they do not affect the amino acid sequence of the protein (Achtman *et al.*, 1999; Alm *et al.*, 1999; Suerbaum *et al.*, 1998). About 30 genes have been found to be associated with stretches of mono-or dinucleotide repeats, which are liable to slipped strand mispairing that can result in translational frameshifts and stop codons, offering means for regulation of gene expression (Alm *et al.*, 1999; Salaün *et al.*, 2005; Tomb *et al.*, 1997). These phase-variable genes commonly encode proteins involved in restriction-modification, lipopolysaccharide (LPS) biosynthesis and cell surface-associated proteins.

3.3. Virulence factors

A virulence factor contributes some function that renders the microorganism more pathogenic, that is, increases the likelihood for disease development. *H. pylori* infection is usually lifelong and asymptomatic and disease may be attributed to the host response towards the colonization. Thus, some of the factors commonly designated as virulence factors in *H. pylori*, for instance the flagella, may rather be regarded as "colonization factors" (Testerman *et al.*, 2001). These factors primarily facilitate establishment and persistence of the infection, which however, naturally also increases the risk for disease, blurring a distinction from virulence factors. Studies of the contributions of individual bacterial factors to infectivity and pathogenicity have to be cautiously interpreted. Associations between different factors may give rise to confounding (Atherton *et al.*, 1995; Gerhard *et al.*, 1999; Xiang *et al.*, 1995; Zambon *et al.*, 2003) and the influences of unrecognized subtle mutations may result in spurious findings (Salaün *et al.*, 2005; Yamaoka *et al.*, 2002).

The cag PAI and VacA

The cag PAI is one of the most studied loci in the H. pylori genome and is present in the majority of strains worldwide (Nilsson et al., 2003; Park et al., 2002; Parsonnet et al., 1997; Simán et al., 2005). The locus is associated with a more vigorous host response characterized by IL-8 induction (Akopyants et al., 1998; Censini et al., 1996; Nilsson et al., 2003; Yamaoka et al., 1996) and an increased risk for ulceration and cancer (Gerhard et al., 1999; Nilsson et al., 2003; Nomura et al., 2002; Parsonnet et al., 1997; Zambon et al., 2003). The cag PAI is an almost 40 kb stretch of DNA that encodes nearly 30 genes, many of which are homologous to type IV secretion system components (Akopyants et al., 1998; Censini et al., 1996; Odenbreit et al., 2000). Type IV secretion systems assemble into a syringe-like structure that mediates secretion of molecules extracellularly or into the cytosol of host cells. The secretion system of H. pylori delivers the cag PAI-encoded and immunodominant CagA protein into the gastric epithelial cells (Odenbreit et al., 2000; Segal et al., 1999). Upon translocation, CagA is phosphorylated and initiates signal transduction that results in cytoskeletal rearrangements and an inflammatory response (Brandt et al., 2005; Higashi et al., 2002; Segal et al., 1999). The secretion system may also mediate transfer of *H. pylori* peptidoglycan into the epithelial cells where Nod1, an intracellular pathogen-recognition molecule, can initiate an immune response (Viala et al., 2004). PAIs are typically prone to horizontal genetic transfer. The cag PAI exhibits signs of such mobility by the differing GC content compared to the rest of the genome and the presence of flanking direct repeats and insertion sequences (Akopyants et al., 1998; Censini et al., 1996). Accordingly, excision and insertion of the cag PAI can result in mixed infections with regard to cag PAI status (Björkholm et al., 2001a; Kersulyte et al., 1999; Tomasini et al., 2003). Intermediate strains that lack some of the cag PAI genes have also been described (Nilsson et al., 2003; Tomasini et al., 2003).

Early on, *H. pylori* was found to possess a cytotoxic ability involving formation of vacuoles in epithelial cells, which could be attributed to the bacterial exotoxin VacA (Cover and Blaser, 1992). The *vacA* gene appears to be universally present, but there are alleles with different signal (s1/s2) and mid regions (m1/m2) (Atherton *et al.*, 1995). The s1/m1 variant is most cytotoxic and the s1 and m1 genotypes have been proposed to be correlated with the pathogenic potential of the infection (Atherton *et al.*, 1995; Gerhard *et al.*, 1999; Zambon *et al.*, 2003). VacA can induce apoptosis of gastric cells, which may provide the bacteria with nutrients or reduce the acid output through the killing of parietal cells (Boquet *et al.*, 2003; Cover *et al.*, 2003). Furthermore, VacA-mediated inhibition of antigen presentation (Molinari *et al.*, 1998) and activation of T lymphocytes could play a role in immune evasion (Boncristiano *et al.*, 2003; Gebert *et al.*, 2003).

The cytotoxic variant of vacA is in linkage disequilibrium with the *cag* PAI (Atherton *et al.*, 1995; Xiang *et al.*, 1995), hence the gene name *cytotoxin-associated* gene A (*cagA*). The *vacA* and *cag* PAI loci are situated at distant sites on the chromosome and their linkage is inadequately understood. Nevertheless, the loci form the basis for a classification of virulence of *H. pylori* strains. The more virulent

type I strains express CagA and a cytotoxic variant of VacA, while the less virulent type II strains do not express CagA and harbor a non-toxic form of VacA (Xiang *et al.*, 1995). The serological response against CagA has been used as a marker of more virulent strains, but serological methods have been questioned due to limited sensitivity (0.71–0.90) and specificity (0.80–0.90) (Figueiredo *et al.*, 2001; Park *et al.*, 2002; Yamaoka *et al.*, 1998).

3.4. Surface structures

Flagella

The unipolar flagella of *H. pylori* enable motility, which is an important bacterial feature (Eaton *et al.*, 1992; Terry *et al.*, 2005). Two different flagellin proteins constitute the flagellar filament, but about 40 additional genes are involved in the secretion and assembly of the whole flagellar apparatus (Tomb *et al.*, 1997). Chemotactic systems offer means for spatial orientation, for example towards the mucosal cell lining where the pH is higher, nutrients can be more abundant and closer interactions with host cells are possible (Terry *et al.*, 2005).

Outer membrane proteins

A relatively large proportion (4%) of the coding capacity of the *H. pylori* genome is devoted to outer membrane proteins (OMP) (Doig et al., 1999). Several of these proteins have been suggested to possess adhesive properties. The receptors include glycoconjugates expressed on host cells, such as the Lewis carbohydrate blood group antigens, extracellular matrix components and unknowns (Testerman et al., 2001). Two OMPs have received particular attention for their ability to bind to host receptors. First, the BabA adhesin binds to Lewis b that is expressed by gastric epithelial cells (Ilver et al., 1998) and the presence of this adhesin has been suggested to be associated with more severe disease (Gerhard et al., 1999; Zambon et al., 2003). Second, the SabA adhesin mediates a weaker and more intimate adherence by binding to sialyl-Lewis x, which is upregulated by the inflammation (Mahdavi et al., 2002). Accordingly, a model was proposed where initial binding is mediated by Lewis b and BabA, which results in inflammation, induction of sialyl-Lewis x and binding through SabA. Altered adhesive properties may provide mechanisms for *H. pylori* to regulate its interactions with the host if, for example, the immune response would necessitate less tight adherence (Mahdavi et al., 2002) or when the availability of receptors differ between human populations (Aspholm-Hurtig et al., 2004). Such modulation of the binding properties may occur by changed expression or evolution of functional variants through frameshift mutation or recombination between homologous loci (Ilver et al., 1998; Mahdavi et al., 2002). For instance, the BabA-encoding gene *babA2* is similar to the *babA1* and *babB* genes and recombination events between these loci have been described (Bäckström et al., 2004; Solnick et al., 2004).

Lipopolysaccharide

LPS covers the surface of H. pylori and other gram-negative bacteria and sustains membrane integrity and can mediate interaction with the host. An unusual feature of H. pylori LPS is the expression of Lewis antigens on the polymeric carbohydrate Oantigen constituent, resembling blood group antigens expressed on various host tissues. About 80-90% of H. pylori strains express Lewis antigens, whereof Lewis x and Lewis y are most common (Simoons-Smit et al., 1996; Taylor et al., 1998). Fucosyltransferases involved in the synthesis of Lewis antigens can undergo slipped strand mispairing, generating variability of Lewis antigen expression, which may aid adaptive evolution (Appelmelk et al., 1999; Salaün et al., 2005; Wirth et al., 1999). The molecular mimicry of *H. pylori* and human Lewis antigen expression has been suggested to facilitate bacterial immune evasion (Wirth et al., 1997) and give rise to autoimmunity (Heneghan et al., 2001). H. pylori Lewis antigens could further interact with dendritic cells to balance the Th1/Th2 responses (Bergman et al., 2004) and have been described to mediate adhesion (Edwards et al., 2000). However, much of the evidence regarding the biological roles of H. pylori Lewis antigen expression is inconclusive and awaits confirmatory data (Appelmelk et al., 2000; Mahdavi et al., 2003; Taylor et al., 1998).

4. H. PYLORI-ASSOCIATED DISEASE

The ability of *H. pylori* to maintain persistent colonization is of key importance for disease development. The infection is accompanied by a usually asymptomatic chronic gastritis, but 10–20% of infected individuals manifest more severe complications, such as peptic ulcer disease and gastric cancer (Suerbaum and Michetti, 2002). This selectivity in disease progression is inadequately understood and explanations are sought among factors of the host (Correa *et al.*, 2004; El-Omar *et al.*, 2000), environment (Correa *et al.*, 2004) and bacterium (Gerhard *et al.*, 1999; Nomura *et al.*, 2002; Parsonnet *et al.*, 1997; Zambon *et al.*, 2003).

4.1. Gastritis

Acute *H. pylori* infection causes gastritis and hypochlorhydria and symptoms such as vomiting and dyspepsia have been associated with acquisition (Graham *et al.*, 2004; Marshall, 2001). Persistent infection causes chronic gastritis in virtually all infected individuals. The gastric inflammation involves infiltration of immune cells, such as neutrophils, lymphocytes, plasma cells and macrophages, and secretion of a multitude of cytokines, of which IL-8 seems to have a central role (Blaser and Atherton, 2004; Suerbaum and Michetti, 2002; Yamaoka *et al.*, 1996). The chronic gastritis is usually asymptomatic, but eradication of *H. pylori* in non-ulcer dyspeptic patients alleviates symptoms in a fraction of the patients (Moayyedi *et al.*, 2005).

The chronic gastritis leads to peptic ulcer disease and gastric cancer in 10-20% of infected individuals (Suerbaum and Michetti, 2002). Duodenal ulcer disease and gastric ulcer/cancer represent different pathways of the infection and correlate strongly with the pattern of colonization and gastritis. Duodenal ulceration is characterized by higher acid secretion and antrum-predominant gastritis, while gastric ulceration/carcinogenesis is associated with lower acid secretion and corpus-predominant gastritis or pangastritis (Blaser and Atherton, 2004; Suerbaum and Michetti, 2002; Uemura *et al.*, 2001).

4.2. Peptic ulcer disease

The discovery of the role of *H. pylori* in the development of peptic ulcer disease has lead to a paradigm shift in the treatment of ulcer patients (NIH Consensus Conference, 1994). The lifetime risk for peptic ulcer in infected individuals ranges from 3% in the United States to 25% in Japan (Suerbaum and Michetti, 2002). It has been estimated that 95% of duodenal ulcers and 70% of gastric ulcers can be attributed to *H. pylori* (Rothenbacher and Brenner, 2003).

Duodenal ulcers are associated with *H. pylori*-induced antrum-predominant gastritis, decreased somatostatin levels and augmented gastrin and acid secretion (Blaser and

Atherton, 2004; Suerbaum and Michetti, 2002). Development of gastric metaplasia in the duodenum can allow further bacterial colonization, leading to duodenitis and epithelial damage. Gastric ulcers are associated with corpus gastritis, which is believed to damage the epithelium (Blaser and Atherton, 2004). Eradication of the infection heals peptic ulcer disease, restores normal acid secretion and prevents ulcer relapse (Ford *et al.*, 2004).

4.3. Gastric cancer

Gastric cancer ranks as the second most frequent cause of cancer deaths worldwide despite a decreasing incidence in high-income countries (Correa *et al.*, 2004). The World Health Organization classified *H. pylori* as a class I carcinogen in 1994 due to its definite carcinogenic potential in humans (IARC, 1994). Subsequent studies have corroborated the association to gastric cancer and about 70% of non-cardia adenocarcinomas have been attributed to *H. pylori* infection (Ekström *et al.*, 2001; Uemura *et al.*, 2001). However, only a few percent of infected individuals develop gastric cancer (Blaser and Atherton, 2004; Uemura *et al.*, 2001). *H. pylori* acquisition in younger ages has been suggested to contribute a greater cancer risk (Blaser *et al.*, 1995), but this study is limited by its sparse adjustments and its use of sibship size and birth order as proxies for early acquisition.

Persons with low gastric acid secretion and corpus-predominant gastritis, leading to atrophic gastritis, loss of parietal cells and further hypochlorhydria, are at increased risk for gastric cancer (Blaser and Atherton, 2004; Suerbaum and Michetti, 2002; Uemura *et al.*, 2001). Postulated carcinogenic mechanisms include the increased epithelial turnover caused by the inflammation. Furthermore, the development of gastric atrophy and hypochlorhydria can result in impaired antioxidant absorption, infection with other carcinogenic microorganisms and formation of carcinogenic compounds (Blaser and Atherton, 2004). *H. pylori* infection upregulates the pro-inflammatory cytokine IL-1 β , which is also a potent inhibitor of acid secretion. Polymorphisms considered to increase the activity of IL-1 β have been proposed to be associated with hypochlorhydria and cancer, supporting the central role of gastric acidity in cancer development (El-Omar *et al.*, 2000). The atrophic stomach appears to constitute an inhospitable environment for *H. pylori* and cleared infections can lead to misclassification of exposure and underestimation of the gastric cancer risk associated with the infection (Blaser and Atherton, 2004; Ekström *et al.*, 2001).

4.4. Other H. pylori-associated conditions

H. pylori has been assessed for its involvement in various additional conditions. An association between the infection and gastric mucosa-associated lymphoid tissue (MALT) lymphoma belongs among the generally accepted findings and antibiotic treatment alone leads to regression of the cancer in many cases (Farinha and Gascoyne, 2005). Moreover, it has been debated whether there is a causal

relationship between the parallel decline of *H. pylori* prevalence and the increase of gastroesophageal reflux disease (GERD) and esophageal adenocarcinoma in highincome countries. There is accumulating evidence in favor of a protective role of *H. pylori* infection against these conditions, but there are a number of additional contributing factors (Lagergren, 2005; Malfertheiner and Peitz, 2005).

The high *H. pylori* prevalence in many parts of the world accentuates the public health importance of any associations between *H. pylori* and disease. This may be particularly noteworthy for associations that are of modest strength or involve relatively benign conditions, as these associations may otherwise tend to be neglected. In this context, a possible role of the infection in malnutrition and iron deficiency anemia may be of special interest (Milman *et al.*, 1998; Salgueiro *et al.*, 2004), given the high *H. pylori* prevalence in low-income countries where malnutrition can be common for other reasons. *H. pylori* infection has, however, also been suggested to provide protection against childhood diarrheal disease (Rothenbacher *et al.*, 2000) and *H. pylori*-induced anemia has been hypothesized to protect against malaria (Dominguez-Bello and Blaser, 2005).

5. MANAGEMENT OF H. PYLORI INFECTION

Clinical management of *H. pylori* infection and its associated morbidity, as well as related research, rely on the availability of practical and well-evaluated diagnostic tests and treatments. Disease prevention may be possible by targeting the infection, either by eradication treatment or by preventing the establishment of the infection. Furthermore, it should be kept in mind that *H. pylori* infection is most prevalent in low-income countries and any tools for the management of the infection should thus preferably be accessible also in these countries.

5.1. Diagnosis

H. pylori can be detected through endoscopy by culture, histology or urease test of biopsies (Malfertheiner *et al.*, 2002; Suerbaum and Michetti, 2002). These methods are all liable to false negative results due to patchy bacterial colonization. Culture is the theoretical gold standard for identifying bacterial infections, but can have low sensitivity for *H. pylori*.

The infection elicits a systemic IgG antibody response that can be detected for diagnostic purposes (Malfertheiner *et al.*, 2002; Suerbaum and Michetti, 2002). Anti-*H. pylori* antibodies can be assessed with enzyme-linked immunosorbent assays (ELISA) or Western Blot (also designated immunoblot). The latter method has the advantage of characterizing the immune response towards different bacterial antigens. Serology can have limited sensitivity in young children and the antigenic preparation may influence the results (Malfertheiner *et al.*, 2002; Suerbaum and Michetti, 2002; Tindberg *et al.*, 2001a).

The ¹³C-urea breath test (UBT) relies on the principle that ¹³C-labeled urea is hydrolyzed by the bacterial urease with formation of ¹³CO₂, which is detected in the expired breath (Malfertheiner *et al.*, 2002; Suerbaum and Michetti, 2002). An advantage of the UBT is its ability to detect current infection, but the performance of the test in the youngest children has been questioned. There are also reliable ELISAbased stool antigen tests that detect *H. pylori* antigens shed in the feces. Current guidelines recommend the UBT or the stool antigen test for diagnosing the infection in primary care because serological methods require local validation (Malfertheiner *et al.*, 2002). The UBT or, if the UBT is not available, the stool antigen test are recommended to confirm eradication of infection after treatment.

5.2. Treatment

The relatively benign nature of *H. pylori* infection in the majority of infected individuals has elicited debate about how a positive diagnostic test should be handled. The argumentation has been further fueled by the suggested protective

effect of the infection on esophageal adenocarcinoma. Nevertheless, general guidelines for treatment of the infection have been developed and continue to evolve (Malfertheiner *et al.*, 2002; Suerbaum and Michetti, 2002). *H. pylori* eradication is strongly recommended for all patients with peptic ulcer disease, MALT lymphoma, atrophic gastritis, after gastric cancer resection and for first-degree relatives of gastric cancer patients. Furthermore, eradication treatment is advised for patients taking non-steroidal anti-inflammatory drugs (NSAID), which is an independent risk factor for peptic ulcer disease, and for GERD patients on long-term profound acid suppression, who can develop corpus atrophic gastritis. Patients presenting with persistent dyspepsia may also be offered eradication treatment, as it may lead to symptom improvement in a subset of the cases.

H. pylori eradication treatment lasts for one to two weeks and usually includes two antimicrobials and an antisecretory agent because acid impairs the efficiency of some antibiotics (Malfertheiner *et al.*, 2002; Suerbaum and Michetti, 2002). The principal antimicrobials are clarithromycin, amoxicillin, metronidazole and tetracycline and the acid suppressant is usually a proton pump inhibitor. Ranitidine bismuth citrate combines antibacterial and antisecretory activities and can also be used. The cure rate is 80% or above, but antibiotic resistance to particularly metronidazole and clarithromycin is an increasing concern (Malfertheiner *et al.*, 2002; Suerbaum and Michetti, 2002).

Reinfection rates are generally considered to be low after successful eradication (Farrell *et al.*, 2004; Mitchell *et al.*, 1998; Rowland *et al.*, 1999; Suerbaum and Michetti, 2002). However, reinfection may be more common in young children (Magistà *et al.*, 2005; Rowland *et al.*, 1999) and in high-prevalence settings (Hildebrand *et al.*, 2001; Soto *et al.*, 2003; Wheeldon *et al.*, 2005). Post-eradication reinfection rates of about 20% have been reported in adults in high-prevalence communities (Soto *et al.*, 2003; Wheeldon *et al.*, 2005), thus being comparable to the incidence in childhood. These reported high reinfection rates speak against a significant role of protective immunity after therapeutic eradication and indicate that prevention of acquisition is needed to attain long-term absence of infection in some high-prevalence settings.

5.3. Prevention of H. pylori-associated disease

Prevention of *H. pylori*-associated disease benefits from predictions of who will become clinically ill. Accordingly, current treatment guidelines advise prophylactic *H. pylori* eradication for some individuals at higher risk for disease, for example patients with atrophic gastritis or taking NSAIDs (Malfertheiner *et al.*, 2002; Suerbaum and Michetti, 2002). Some studies have also targeted high-risk population groups to study the effect of *H. pylori* eradication. Anti-*H. pylori* treatment has been reported to increase regression of cancer precursor lesions (Correa *et al.*, 2000; Kuipers *et al.*, 2004) and, despite low power and a lack of studies, there is some evidence that *H. pylori* eradication may protect against gastric cancer (Uemura *et al.*,

2001; Wong *et al.*, 2004). Future prevention approaches may possibly benefit from a deeper knowledge of the pathogenic mechanisms by allowing more precise identification of individuals at high risk for disease.

Indiscriminate treatment of *H. pylori* infections has been proposed as an approach to limit the burden of *H. pylori*-associated disease. The appropriateness of such a large-scale and crude intervention has been questioned due to the uncertain full spectrum of possible harmful consequences, for example the development of antibiotic resistance (Malfertheiner *et al.*, 2002; Sjölund *et al.*, 2003; Suerbaum and Michetti, 2002). Testing and treating large numbers of persons would also imply significant costs and would therefore be unrealistic in many parts of the world.

An alternative approach could be to target the acquisition or persistence of the infection, while limiting the use of antibiotics. The role of an *H. pylori* vaccine is uncertain given the common failure of the immune system to clear the infection and the apparently inadequate protective immunity against reinfection (Hildebrand *et al.*, 2001; Magistà *et al.*, 2005; Soto *et al.*, 2003; Wheeldon *et al.*, 2005). A protective vaccine would also have to be administered at an early age before the infection is acquired. At this age, an immature immune system may not respond sufficiently to immunization. Another approach could perhaps be a therapeutic vaccine that would circumvent problems with antibiotic resistance. There have been considerable efforts to develop vaccines against *H. pylori*, but despite some encouraging results further work is needed to bring about effective and safe candidates for humans (Ruggiero *et al.*, 2003). Moreover, probiotics have been suggested to be capable of contributing to control of *H. pylori* infection, but this area of research is in its infancy (Hamilton-Miller, 2003).

Preventing establishment of infection by interfering with transmission is a strategy that has been used in public health interventions against a variety of infections. Only a few smaller trials have considered preventing the establishment of *H. pylori* infection by limiting the transmission. This can partly be explained by the fact that there is no apparent prevention strategy at present. The lack of thinkable interventions may be attributed to the seemingly multifaceted nature of *H. pylori* acquisition, intertwined with activities of everyday life. One study reported that the introduction of a lidded, narrow-mouthed water vessel into households was protective against seroconversion (Glynn *et al.*, 2002). In the same study, however, fecal contamination of the water source and using a water disinfectant were not related to becoming infected. There has also been an attempt to detect a difference in the reinfection rates in children depending on whether the whole family unit received eradication therapy or not (Farrell *et al.*, 2004). No such difference was detected, but the authors acknowledged that the study was likely underpowered due to an overall low reinfection rate.

Any future prevention of *H. pylori*-associated disease should likely be primarily aimed at high-risk populations and target both the infection and other known risk factors. Antibiotic treatment is likely to play a central role in efforts to eliminate the

infection. However, understanding and interfering with the acquisition or persistence of the infection by other means may become useful supplemental strategies. This is likely to be especially true in some (low-income) populations, where effective antibiotic regimens may be impaired by high cost, poor compliance, antibiotic resistance and high reinfection rates.

6. AIMS OF THE PRESENT INVESTIGATION

The general aim of this thesis was to advance our understanding of some aspects of *H. pylori* infection, specifically related to its transmission, and highlight some methodological issues.

Specific aims were to:

- Disentangle the independent contributions of *H. pylori* infections in mothers, fathers and siblings to the risk for the infection in 11- to 13-year-old index children (Paper I).
- Investigate if appropriate analysis of all available data, including additional non-randomly sampled cases, would reveal further insights into the familial clustering of *H. pylori* infection, while possibly improving the precision of the estimates of risk factors already identified (Paper II).
- Explore patterns of *H. pylori* strain concordance in bacterial isolates from family members by using PCR-based molecular typing (Paper III).
- Assess the suitability of serology for determining the *cag* PAI status of *H. pylori* infections, by investigating the agreement between the occurrence of CagA-reactive antibodies, as assessed by immunoblot, and bacterial *cag* PAI status, as determined by PCR (Paper IV).
- Explore the genetic diversity of clonal *H. pylori* isolates within and between members of a family by sequencing and comparative genomic microarray hybridizations (Paper V).

The following sections summarize the methods and results of Papers I–V and bring the findings together in a context of the current literature. More detailed information regarding the individual studies can be retrieved from the corresponding papers that are provided as appendices.

7. SUBJECTS AND METHODS

Sections 7, 8, 9 and 13 are omitted from the electronic version of the thesis due to copyright reasons. Please refer to the original papers.

8. RESULTS

Sections 7, 8, 9 and 13 are omitted from the electronic version of the thesis due to copyright reasons. Please refer to the original papers.

9. DISCUSSION

Sections 7, 8, 9 and 13 are omitted from the electronic version of the thesis due to copyright reasons. Please refer to the original papers.

10. CONCLUDING REMARKS

Infectious diseases and their sequelae are by definition preventable. In this respect, understanding and interfering with the transmission, colonization or pathogenesis of the infectious agent are fundamental. There are some promising data on prevention of *H. pylori*-associated morbidity by treating the infection in high-risk populations, but attempts to limit the transmission are lacking.

The identification of the family and early childhood as the primary place and time for *H. pylori* acquisition are important pieces of epidemiological knowledge. The present data support a predominantly mother-child and sib-sib transmission of *H. pylori*. This finding is consistent with an important role of intimate contact in the transmission and a relatively low infectiousness of the bacterium. The extensive *H. pylori* genome variability is considered to be important for adaptive evolution and we discerned characteristics of the variability likely to be important for the bacterium. This diversity could, however, not be related to either transmission or host adaptation. Further epidemiological and microbiological insights into *H. pylori* acquisition, persistence and pathogenesis may improve clinical and public health management of the infection. Thus, these issues deserve sustained attention and longitudinal studies of human populations coupled with collection of bacterial samples and experimental investigations could be valuable undertakings. The findings presented in this thesis and the discussions thereof may aid in directing such future endeavors.

For any public health intervention to be ethically defendable, benefits and possible detrimental side effects must be carefully evaluated beforehand. A parallel is found in the study of hazardous exposures in humans, which does not allow randomized experimental approaches. Epidemiology has solved this predicament by providing tools for appropriate analyses of data from the "natural experiment", that is, a population where the exposure is already present. Social and economic development results in a declining *H. pylori* prevalence, not to mention the many other more tangible benefits. However, the specific factors that cause this decreasing prevalence, as well as its consequences for humans, remain elusive. Part of the solution to a better understanding of the relationship between humans and *H. pylori* perhaps lies within clever exploration of this already operative "social vaccine" against *H. pylori* infection.

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12. REFERENCES

- Achtman, M., Azuma, T., Berg, D.E., Ito, Y., Morelli, G., Pan, Z.J., Suerbaum, S., Thompson, S.A., van der Ende, A., and van Doorn, L.J. (1999) Recombination and clonal groupings within *Helicobacter pylori* from different geographical regions. *Mol Microbiol* 32: 459-470.
- Akopyants, N.S., Clifton, S.W., Kersulyte, D., Crabtree, J.E., Youree, B.E., Reece, C.A., Bukanov, N.O., Drazek, E.S., Roe, B.A., and Berg, D.E. (1998) Analyses of the *cag* pathogenicity island of *Helicobacter pylori*. *Mol Microbiol* 28: 37-53.
- Akopyanz, N., Bukanov, N.O., Westblom, T.U., Kresovich, S., and Berg, D.E. (1992) DNA diversity among clinical isolates of *Helicobacter pylori* detected by PCR-based RAPD fingerprinting. *Nucleic Acids Res* 20: 5137-5142.
- Allen, L.A. (2001) The role of the neutrophil and phagocytosis in infection caused by *Helicobacter pylori. Curr Opin Infect Dis* 14: 273-277.
- Alm, R.A., Ling, L.S., Moir, D.T., King, B.L., Brown, E.D., Doig, P.C., Smith, D.R., Noonan, B., Guild, B.C., deJonge, B.L., Carmel, G., Tummino, P.J., Caruso, A., Uria-Nickelsen, M., Mills, D.M., Ives, C., Gibson, R., Merberg, D., Mills, S.D., Jiang, Q., Taylor, D.E., Vovis, G.F., and Trust, T.J. (1999) Genomicsequence comparison of two unrelated isolates of the human gastric pathogen *Helicobacter pylori. Nature* 397: 176-180.
- Andersen, L.P., Dorland, A., Karacan, H., Colding, H., Nilsson, H.O., Wadström, T., and Blom, J. (2000) Possible clinical importance of the transformation of *Helicobacter pylori* into coccoid forms. *Scand J Gastroenterol* 35: 897-903.
- Andersen-Nissen, E., Smith, K.D., Strobe, K.L., Barrett, S.L., Cookson, B.T., Logan, S.M., and Aderem, A. (2005) Evasion of Toll-like receptor 5 by flagellated bacteria. *Proc Natl Acad Sci U S A* 102: 9247-9252.
- Appelmelk, B.J., Martin, S.L., Monteiro, M.A., Clayton, C.A., McColm, A.A., Zheng, P., Verboom, T., Maaskant, J.J., van den Eijnden, D.H., Hokke, C.H., Perry, M.B., Vandenbroucke-Grauls, C.M., and Kusters, J.G. (1999) Phase variation in *Helicobacter pylori* lipopolysaccharide due to changes in the lengths of poly(C) tracts in alpha3-fucosyltransferase genes. *Infect Immun* 67: 5361-5366.
- Appelmelk, B.J., Monteiro, M.A., Martin, S.L., Moran, A.P., and Vandenbroucke-Grauls, C.M. (2000) Why *Helicobacter pylori* has Lewis antigens. *Trends Microbiol* 8: 565-570.
- Aspholm-Hurtig, M., Dailide, G., Lahmann, M., Kalia, A., Ilver, D., Roche, N., Vikström, S., Sjöström, R., Lindén, S., Bäckström, A., Lundberg, C., Arnqvist, A., Mahdavi, J., Nilsson, U.J., Velapatino, B., Gilman, R.H., Gerhard, M., Alarcon, T., Lopez-Brea, M., Nakazawa, T., Fox, J.G., Correa, P., Dominguez-Bello, M.G., Perez-Perez, G.I., Blaser, M.J., Normark, S., Carlstedt, I., Oscarson, S., Teneberg, S., Berg, D.E., and Borén, T. (2004) Functional adaptation of BabA, the *H. pylori* ABO blood group antigen binding adhesin. *Science* 305: 519-522.

- Atherton, J.C., Cao, P., Peek, R.M., Jr., Tummuru, M.K., Blaser, M.J., and Cover, T.L. (1995) Mosaicism in vacuolating cytotoxin alleles of *Helicobacter pylori*. Association of specific *vacA* types with cytotoxin production and peptic ulceration. *J Biol Chem* 270: 17771-17777.
- Azuma, T., Konishi, J., Ito, Y., Hirai, M., Tanaka, Y., Ito, S., Kato, T., and Kohli, Y. (1995) Genetic differences between duodenal ulcer patients who were positive or negative for *Helicobacter pylori*. J Clin Gastroenterol 21 Suppl 1: S151-154.
- Bamford, K.B., Bickley, J., Collins, J.S., Johnston, B.T., Potts, S., Boston, V., Owen, R.J., and Sloan, J.M. (1993) *Helicobacter pylori*: Comparison of DNA fingerprints provides evidence for intrafamilial infection. *Gut* 34: 1348-1350.
- Banatvala, N., Mayo, K., Megraud, F., Jennings, R., Deeks, J.J., and Feldman, R.A. (1993) The cohort effect and *Helicobacter pylori*. J Infect Dis 168: 219-221.
- Bayerdörffer, E., Oertel, H., Lehn, N., Kasper, G., Mannes, G.A., Sauerbruch, T., and Stolte, M. (1989) Topographic association between active gastritis and *Campylobacter pylori* colonisation. *J Clin Pathol* 42: 834-839.
- Berg, D.E., Gilman, R.H., Lelwala-Guruge, J., Srivastava, K., Valdez, Y., Watanabe, J., Miyagi, J., Akopyants, N.S., Ramirez-Ramos, A., Yoshiwara, T.H., Recavarren, S., and Leon-Barua, R. (1997) *Helicobacter pylori* populations in Peruvian patients. *Clin Infect Dis* 25: 996-1002.
- Bergenzaun, P., Kristinsson, K.G., Thjodleifsson, B., Sigvaldadottir, E., Mölstad, S., Held, M., and Wadström, T. (1996) Seroprevalence of *Helicobacter pylori* in south Sweden and Iceland. *Scand J Gastroenterol* 31: 1157-1161.
- Bergman, M.P., Engering, A., Smits, H.H., van Vliet, S.J., van Bodegraven, A.A., Wirth, H.P., Kapsenberg, M.L., Vandenbroucke-Grauls, C.M., van Kooyk, Y., and Appelmelk, B.J. (2004) *Helicobacter pylori* modulates the T helper cell 1/T helper cell 2 balance through phase-variable interaction between lipopolysaccharide and DC-SIGN. *J Exp Med* 200: 979-990.
- Björkholm, B., Zhukhovitsky, V., Löfman, C., Hultén, K., Enroth, H., Block, M., Rigo, R., Falk, P., and Engstrand, L. (2000) *Helicobacter pylori* entry into human gastric epithelial cells: A potential determinant of virulence, persistence, and treatment failures. *Helicobacter* 5: 148-154.
- Björkholm, B., Lundin, A., Sillén, A., Guillemin, K., Salama, N., Rubio, C., Gordon, J.I., Falk, P., and Engstrand, L. (2001a) Comparison of genetic divergence and fitness between two subclones of *Helicobacter pylori*. *Infect Immun* 69: 7832-7838.
- Björkholm, B., Sjölund, M., Falk, P.G., Berg, O.G., Engstrand, L., and Andersson, D.I. (2001b) Mutation frequency and biological cost of antibiotic resistance in *Helicobacter pylori*. *Proc Natl Acad Sci U S A* 98: 14607-14612.
- Björkholm, B., Guruge, J., Karlsson, M., O'Donnell, D., Engstrand, L., Falk, P., and Gordon, J. (2004) Gnotobiotic transgenic mice reveal that transmission of *Helicobacter pylori* is facilitated by loss of acid-producing parietal cells in donors and recipients. *Microbes Infect* 6: 213-220.
- Blaser, M.J., Chyou, P.H., and Nomura, A. (1995) Age at establishment of *Helicobacter pylori* infection and gastric carcinoma, gastric ulcer, and duodenal ulcer risk. *Cancer Res* 55: 562-565.

- Blaser, M.J., and Berg, D.E. (2001) *Helicobacter pylori* genetic diversity and risk of human disease. *J Clin Invest* 107: 767-773.
- Blaser, M.J., and Atherton, J.C. (2004) *Helicobacter pylori* persistence: Biology and disease. J Clin Invest 113: 321-333.
- Blecker, U., Lanciers, S., Keppens, E., and Vandenplas, Y. (1994) Evolution of *Helicobacter pylori* positivity in infants born from positive mothers. *J Pediatr Gastroenterol Nutr* 19: 87-90.
- Boncristiano, M., Paccani, S.R., Barone, S., Ulivieri, C., Patrussi, L., Ilver, D., Amedei, A., D'Elios, M.M., Telford, J.L., and Baldari, C.T. (2003) The *Helicobacter pylori* vacuolating toxin inhibits T cell activation by two independent mechanisms. *J Exp Med* 198: 1887-1897.
- Boneca, I.G., de Reuse, H., Epinat, J.C., Pupin, M., Labigne, A., and Moszer, I. (2003) A revised annotation and comparative analysis of *Helicobacter pylori* genomes. *Nucleic Acids Res* 31: 1704-1714.
- Boquet, P., Ricci, V., Galmiche, A., and Gauthier, N.C. (2003) Gastric cell apoptosis and *H. pylori*: Has the main function of VacA finally been identified? *Trends Microbiol* 11: 410-413.
- Borén, T., Falk, P., Roth, K.A., Larson, G., and Normark, S. (1993) Attachment of *Helicobacter pylori* to human gastric epithelium mediated by blood group antigens. *Science* 262: 1892-1895.
- Brandt, S., Kwok, T., Hartig, R., König, W., and Backert, S. (2005) NF-kappaB activation and potentiation of proinflammatory responses by the *Helicobacter pylori* CagA protein. *Proc Natl Acad Sci U S A* 102: 9300-9305.
- Brenner, H., Rothenbacher, D., Bode, G., Dieudonne, P., and Adler, G. (1999) Active infection with *Helicobacter pylori* in healthy couples. *Epidemiol Infect* 122: 91-95.
- Brown, L.M., Thomas, T.L., Ma, J.L., Chang, Y.S., You, W.C., Liu, W.D., Zhang, L., Pee, D., and Gail, M.H. (2002) *Helicobacter pylori* infection in rural China: Demographic, lifestyle and environmental factors. *Int J Epidemiol* 31: 638-645.
- Burucoa, C., Lhomme, V., and Fauchere, J.L. (1999) Performance criteria of DNA fingerprinting methods for typing of *Helicobacter pylori* isolates: Experimental results and meta-analysis. *J Clin Microbiol* 37: 4071-4080.
- Bäckhed, F., Rokbi, B., Torstensson, E., Zhao, Y., Nilsson, C., Seguin, D., Normark, S., Buchan, A.M., and Richter-Dahlfors, A. (2003) Gastric mucosal recognition of *Helicobacter pylori* is independent of Toll-like receptor 4. J Infect Dis 187: 829-836.
- Bäckström, A., Lundberg, C., Kersulyte, D., Berg, D.E., Borén, T., and Arnqvist, A. (2004) Metastability of *Helicobacter pylori bab* adhesin genes and dynamics in Lewis b antigen binding. *Proc Natl Acad Sci U S A* 101: 16923-16928.
- Censini, S., Lange, C., Xiang, Z., Crabtree, J.E., Ghiara, P., Borodovsky, M., Rappuoli, R., and Covacci, A. (1996) *cag*, a pathogenicity island of *Helicobacter pylori*, encodes type I-specific and disease-associated virulence factors. *Proc Natl Acad Sci USA* 93: 14648-14653.
- Clemens, J., Albert, M.J., Rao, M., Huda, S., Qadri, F., Van Loon, F.P., Pradhan, B., Naficy, A., and Banik, A. (1996) Sociodemographic, hygienic and nutritional

correlates of *Helicobacter pylori* infection of young Bangladeshi children. *Pediatr Infect Dis J* 15: 1113-1118.

- Clyne, M., and Drumm, B. (1997) Absence of effect of Lewis A and Lewis B expression on adherence of *Helicobacter pylori* to human gastric cells. *Gastroenterology* 113: 72-80.
- Cook, G.C. (1985) Infective gastroenteritis and its relationship to reduced gastric acidity. *Scand J Gastroenterol Suppl* 111: 17-23.
- Correa, P., Fontham, E.T., Bravo, J.C., Bravo, L.E., Ruiz, B., Zarama, G., Realpe, J.L., Malcom, G.T., Li, D., Johnson, W.D., and Mera, R. (2000) Chemoprevention of gastric dysplasia: Randomized trial of antioxidant supplements and anti-*Helicobacter pylori* therapy. J Natl Cancer Inst 92: 1881-1888.
- Correa, P., Piazuelo, M.B., and Camargo, M.C. (2004) The future of gastric cancer prevention. *Gastric Cancer* 7: 9-16.
- Cover, T.L., and Blaser, M.J. (1992) Purification and characterization of the vacuolating toxin from *Helicobacter pylori*. J Biol Chem 267: 10570-10575.
- Cover, T.L., Krishna, U.S., Israel, D.A., and Peek, R.M., Jr. (2003) Induction of gastric epithelial cell apoptosis by *Helicobacter pylori* vacuolating cytotoxin. *Cancer Res* 63: 951-957.
- Doig, P., de Jonge, B.L., Alm, R.A., Brown, E.D., Uria-Nickelsen, M., Noonan, B., Mills, S.D., Tummino, P., Carmel, G., Guild, B.C., Moir, D.T., Vovis, G.F., and Trust, T.J. (1999) *Helicobacter pylori* physiology predicted from genomic comparison of two strains. *Microbiol Mol Biol Rev* 63: 675-707.
- Dominguez-Bello, M.G., and Blaser, M.J. (2005) Are iron-scavenging parasites protective against malaria? *J Infect Dis* 191: 646.
- Dore, M.P., Bilotta, M., Vaira, D., Manca, A., Massarelli, G., Leandro, G., Atzei, A., Pisanu, G., Graham, D.Y., and Realdi, G. (1999) High prevalence of *Helicobacter pylori* infection in shepherds. *Dig Dis Sci* 44: 1161-1164.
- Dore, M.P., Malaty, H.M., Graham, D.Y., Fanciulli, G., Delitala, G., and Realdi, G. (2002) Risk factors associated with *Helicobacter pylori* infection among children in a defined geographic area. *Clin Infect Dis* 35: 240-245.
- Drumm, B., Perez-Perez, G.I., Blaser, M.J., and Sherman, P.M. (1990) Intrafamilial clustering of *Helicobacter pylori* infection. *N Engl J Med* 322: 359-363.
- Dubois, A., Berg, D.E., Incecik, E.T., Fiala, N., Heman-Ackah, L.M., Del Valle, J., Yang, M., Wirth, H.P., Perez-Perez, G.I., and Blaser, M.J. (1999) Host specificity of *Helicobacter pylori* strains and host responses in experimentally challenged nonhuman primates. *Gastroenterology* 116: 90-96.
- Eaton, K.A., Morgan, D.R., and Krakowka, S. (1992) Motility as a factor in the colonisation of gnotobiotic piglets by *Helicobacter pylori*. *J Med Microbiol* 37: 123-127.
- Eaton, K.A., and Krakowka, S. (1994) Effect of gastric pH on urease-dependent colonization of gnotobiotic piglets by *Helicobacter pylori*. *Infect Immun* 62: 3604-3607.
- Edwards, N.J., Monteiro, M.A., Faller, G., Walsh, E.J., Moran, A.P., Roberts, I.S., and High, N.J. (2000) Lewis X structures in the O antigen side-chain promote

adhesion of *Helicobacter pylori* to the gastric epithelium. *Mol Microbiol* 35: 1530-1539.

- Ekström, A.M., Held, M., Hansson, L.E., Engstrand, L., and Nyrén, O. (2001) *Helicobacter pylori* in gastric cancer established by CagA immunoblot as a marker of past infection. *Gastroenterology* 121: 784-791.
- El-Omar, E.M., Carrington, M., Chow, W.H., McColl, K.E., Bream, J.H., Young, H.A., Herrera, J., Lissowska, J., Yuan, C.C., Rothman, N., Lanyon, G., Martin, M., Fraumeni, J.F., Jr., and Rabkin, C.S. (2000) Interleukin-1 polymorphisms associated with increased risk of gastric cancer. *Nature* 404: 398-402.
- Enroth, H., Nyrén, O., and Engstrand, L. (1999) One stomach-one strain: Does *Helicobacter pylori* strain variation influence disease outcome? *Dig Dis Sci* 44: 102-107.
- Falush, D., Kraft, C., Taylor, N.S., Correa, P., Fox, J.G., Achtman, M., and Suerbaum, S. (2001) Recombination and mutation during long-term gastric colonization by *Helicobacter pylori*: Estimates of clock rates, recombination size, and minimal age. *Proc Natl Acad Sci U S A* 98: 15056-15061.
- Falush, D., Wirth, T., Linz, B., Pritchard, J.K., Stephens, M., Kidd, M., Blaser, M.J., Graham, D.Y., Vacher, S., Perez-Perez, G.I., Yamaoka, Y., Megraud, F., Otto, K., Reichard, U., Katzowitsch, E., Wang, X., Achtman, M., and Suerbaum, S. (2003) Traces of human migrations in *Helicobacter pylori* populations. *Science* 299: 1582-1585.
- Farinha, P., and Gascoyne, R.D. (2005) *Helicobacter pylori* and MALT lymphoma. *Gastroenterology* 128: 1579-1605.
- Farrell, S., Milliken, I., Doherty, G.M., Murphy, J.L., Wootton, S.A., and McCallion, W.A. (2004) Total family unit *Helicobacter pylori* eradication and pediatric re-infection rates. *Helicobacter* 9: 285-288.
- Figueiredo, C., Quint, W., Nouhan, N., van den Munckhof, H., Herbrink, P., Scherpenisse, J., de Boer, W., Schneeberger, P., Perez-Perez, G., Blaser, M.J., and van Doorn, L.J. (2001) Assessment of *Helicobacter pylori vacA* and *cagA* genotypes and host serological response. *J Clin Microbiol* 39: 1339-1344.
- Ford, A., Delaney, B., Forman, D., and Moayyedi, P. (2004) Eradication therapy for peptic ulcer disease in *Helicobacter pylori* positive patients. *Cochrane Database Syst Rev*: CD003840.
- Fox, J.G. (2002) The non-*H pylori* helicobacters: Their expanding role in gastrointestinal and systemic diseases. *Gut* 50: 273-283.
- Furuta, T., Kamata, T., Takashima, M., Futami, H., Arai, H., Hanai, H., and Kaneko,
 E. (1997) Study of transmission routes of *Helicobacter pylori* in relation to seroprevalence of hepatitis A virus. *J Clin Microbiol* 35: 1891-1893.
- Gebert, B., Fischer, W., Weiss, E., Hoffmann, R., and Haas, R. (2003) *Helicobacter pylori* vacuolating cytotoxin inhibits T lymphocyte activation. *Science* 301: 1099-1102.
- Gerhard, M., Lehn, N., Neumayer, N., Boren, T., Rad, R., Schepp, W., Miehlke, S., Classen, M., and Prinz, C. (1999) Clinical relevance of the *Helicobacter*

pylori gene for blood-group antigen-binding adhesin. *Proc Natl Acad Sci U S A* 96: 12778-12783.

- Gibson, J.R., Slater, E., Xerry, J., Tompkins, D.S., and Owen, R.J. (1998) Use of an amplified-fragment length polymorphism technique to fingerprint and differentiate isolates of *Helicobacter pylori*. J Clin Microbiol 36: 2580-2585.
- Glynn, M.K., Friedman, C.R., Gold, B.D., Khanna, B., Hutwagner, L., Iihoshi, N., Revollo, C., and Quick, R. (2002) Seroincidence of *Helicobacter pylori* infection in a cohort of rural Bolivian children: Acquisition and analysis of possible risk factors. *Clin Infect Dis* 35: 1059-1065.
- Goodman, K.J., Correa, P., Tengana Aux, H.J., Ramirez, H., DeLany, J.P., Guerrero Pepinosa, O., Lopez Quinones, M., and Collazos Parra, T. (1996) *Helicobacter pylori* infection in the Colombian Andes: A population-based study of transmission pathways. *Am J Epidemiol* 144: 290-299.
- Goodman, K.J., and Correa, P. (2000) Transmission of *Helicobacter pylori* among siblings. *Lancet* 355: 358-362.
- Goodman, K.J., O'Rourke, K., Day, R.S., Wang, C., Nurgalieva, Z., Phillips, C.V., Aragaki, C., Campos, A., and de la Rosa, J.M. (2005) Dynamics of *Helicobacter pylori* infection in a US-Mexico cohort during the first two years of life. *Int J Epidemiol*.
- Goodwin, C.S., Armstrong, J.A., Chilvers, T., Peters, M., Collins, D., Sly, L., McConnel, W., and Harper, W.S. (1989) Transfer of *Campylobacter pylori* and *Campylobacter mustelae* to *Helicobacter* gen. nov. as *Helicobacter pylori* comb. nov. and *Helicobacter mustelae* comb. nov., respectively. *Int J Syst Bacteriol* 4: 397-405.
- Graham, D.Y., Malaty, H.M., Evans, D.G., Evans, D.J., Jr., Klein, P.D., and Adam, E. (1991) Epidemiology of *Helicobacter pylori* in an asymptomatic population in the United States. Effect of age, race, and socioeconomic status. *Gastroenterology* 100: 1495-1501.
- Graham, D.Y., Opekun, A.R., Osato, M.S., El-Zimaity, H.M., Lee, C.K., Yamaoka, Y., Qureshi, W.A., Cadoz, M., and Monath, T.P. (2004) Challenge model for *Helicobacter pylori* infection in human volunteers. *Gut* 53: 1235-1243.
- Granström, M., Tindberg, Y., and Blennow, M. (1997) Seroepidemiology of *Helicobacter pylori* infection in a cohort of children monitored from 6 months to 11 years of age. *J Clin Microbiol* 35: 468-470.
- Greenland, S., and Rothman, K.J. (1998) Modern Epidemiology. Second edition. Measures of effect and measures of association. Rothman, K.J. and Greenland, S. (eds): Lippincott Williams & Wilkins. USA.
- Grübel, P., Hoffman, J.S., Chong, F.K., Burstein, N.A., Mepani, C., and Cave, D.R. (1997) Vector potential of houseflies (*Musca domestica*) for *Helicobacter pylori*. J Clin Microbiol 35: 1300-1303.
- Halloran, M.E. (1998) Modern Epidemiology. Second edition. Concepts of Infectious Disease Epidemiology. Rothman, K.J. and Greenland, S. (eds): Lippincott Williams & Wilkins. USA.
- Hamilton-Miller, J.M. (2003) The role of probiotics in the treatment and prevention of *Helicobacter pylori* infection. *Int J Antimicrob Agents* 22: 360-366.

- Han, S.R., Zschausch, H.C., Meyer, H.G., Schneider, T., Loos, M., Bhakdi, S., and Maeurer, M.J. (2000) *Helicobacter pylori*: Clonal population structure and restricted transmission within families revealed by molecular typing. *J Clin Microbiol* 38: 3646-3651.
- Handt, L.K., Fox, J.G., Dewhirst, F.E., Fraser, G.J., Paster, B.J., Yan, L.L., Rozmiarek, H., Rufo, R., and Stalis, I.H. (1994) *Helicobacter pylori* isolated from the domestic cat: Public health implications. *Infect Immun* 62: 2367-2374.
- Hartland, S., Newton, J.L., Griffin, S.M., and Donaldson, P.T. (2004) A functional polymorphism in the interleukin-1 receptor-1 gene is associated with increased risk of *Helicobacter pylori* infection but not with gastric cancer. *Dig Dis Sci* 49: 1545-1550.
- Hazell, S.L., Mitchell, H.M., Hedges, M., Shi, X., Hu, P.J., Li, Y.Y., Lee, A., and Reiss-Levy, E. (1994) Hepatitis A and evidence against the community dissemination of *Helicobacter pylori* via feces. *J Infect Dis* 170: 686-689.
- Heneghan, M.A., McCarthy, C.F., Janulaityte, D., and Moran, A.P. (2001) Relationship of anti-Lewis x and anti-Lewis y antibodies in serum samples from gastric cancer and chronic gastritis patients to *Helicobacter pylori*mediated autoimmunity. *Infect Immun* 69: 4774-4781.
- Higashi, H., Tsutsumi, R., Muto, S., Sugiyama, T., Azuma, T., Asaka, M., and Hatakeyama, M. (2002) SHP-2 tyrosine phosphatase as an intracellular target of *Helicobacter pylori* CagA protein. *Science* 295: 683-686.
- Hildebrand, P., Bardhan, P., Rossi, L., Parvin, S., Rahman, A., Arefin, M.S., Hasan, M., Ahmad, M.M., Glatz-Krieger, K., Terracciano, L., Bauerfeind, P., Beglinger, C., Gyr, N., and Khan, A.K. (2001) Recrudescence and reinfection with *Helicobacter pylori* after eradication therapy in Bangladeshi adults. *Gastroenterology* 121: 792-798.
- Hopkins, R.J., Vial, P.A., Ferreccio, C., Ovalle, J., Prado, P., Sotomayor, V., Russell, R.G., Wasserman, S.S., and Morris, J.G., Jr. (1993) Seroprevalence of *Helicobacter pylori* in Chile: Vegetables may serve as one route of transmission. *J Infect Dis* 168: 222-226.
- Hultén, K., Han, S.W., Enroth, H., Klein, P.D., Opekun, A.R., Gilman, R.H., Evans, D.G., Engstrand, L., Graham, D.Y., and El-Zaatari, F.A. (1996) *Helicobacter pylori* in the drinking water in Peru. *Gastroenterology* 110: 1031-1035.
- Hunter, P.R., and Gaston, M.A. (1988) Numerical index of the discriminatory ability of typing systems: An application of Simpson's index of diversity. J Clin Microbiol 26: 2465-2466.
- IARC (1994) Schistosomes, liver flukes and *Helicobacter pylori*. IARC monographs on the evaluation of carcinogenic risks to humans. IARC. France.
- Ilver, D., Arnqvist, A., Ögren, J., Frick, I.M., Kersulyte, D., Incecik, E.T., Berg, D.E., Covacci, A., Engstrand, L., and Borén, T. (1998) *Helicobacter pylori* adhesin binding fucosylated histo-blood group antigens revealed by retagging. *Science* 279: 373-377.
- Israel, D.A., Salama, N., Krishna, U., Rieger, U.M., Atherton, J.C., Falkow, S., and Peek, R.M., Jr. (2001) *Helicobacter pylori* genetic diversity within the gastric niche of a single human host. *Proc Natl Acad Sci U S A* 98: 14625-14630.

- Karhukorpi, J., Ikaheimo, I., Silvennoinen-Kassinen, S., Tiilikainen, A.S., and Karttunen, R. (1999) HLA-DQA1 alleles and the presence of *Helicobacter pylori* antibodies. *Eur J Immunogenet* 26: 15-17.
- Kavermann, H., Burns, B.P., Angermuller, K., Odenbreit, S., Fischer, W., Melchers, K., and Haas, R. (2003) Identification and characterization of *Helicobacter pylori* genes essential for gastric colonization. *J Exp Med* 197: 813-822.
- Kersulyte, D., Chalkauskas, H., and Berg, D.E. (1999) Emergence of recombinant strains of *Helicobacter pylori* during human infection. *Mol Microbiol* 31: 31-43.
- Kersulyte, D., Mukhopadhyay, A.K., Velapatino, B., Su, W., Pan, Z., Garcia, C., Hernandez, V., Valdez, Y., Mistry, R.S., Gilman, R.H., Yuan, Y., Gao, H., Alarcon, T., Lopez-Brea, M., Balakrish Nair, G., Chowdhury, A., Datta, S., Shirai, M., Nakazawa, T., Ally, R., Segal, I., Wong, B.C., Lam, S.K., Olfat, F.O., Borén, T., Engstrand, L., Torres, O., Schneider, R., Thomas, J.E., Czinn, S., and Berg, D.E. (2000) Differences in genotypes of *Helicobacter pylori* from different human populations. *J Bacteriol* 182: 3210-3218.
- Klein, P.D., Graham, D.Y., Gaillour, A., Opekun, A.R., and Smith, E.O. (1991) Water source as risk factor for *Helicobacter pylori* infection in Peruvian children. Gastrointestinal physiology working group. *Lancet* 337: 1503-1506.
- Klein, P.D., Gilman, R.H., Leon-Barua, R., Diaz, F., Smith, E.O., and Graham, D.Y. (1994) The epidemiology of *Helicobacter pylori* in Peruvian children between 6 and 30 months of age. *Am J Gastroenterol* 89: 2196-2200.
- Kuepper-Nybelen, J., Thefeld, W., Rothenbacher, D., and Brenner, H. (2005) Patterns of alcohol consumption and *Helicobacter pylori* infection: Results of a population-based study from Germany among 6545 adults. *Aliment Pharmacol Ther* 21: 57-64.
- Kuipers, E.J., Israel, D.A., Kusters, J.G., and Blaser, M.J. (1998) Evidence for a conjugation-like mechanism of DNA transfer in *Helicobacter pylori*. J Bacteriol 180: 2901-2905.
- Kuipers, E.J., Israel, D.A., Kusters, J.G., Gerrits, M.M., Weel, J., van Der Ende, A., van Der Hulst, R.W., Wirth, H.P., Hook-Nikanne, J., Thompson, S.A., and Blaser, M.J. (2000) Quasispecies development of *Helicobacter pylori* observed in paired isolates obtained years apart from the same host. *J Infect Dis* 181: 273-282.
- Kuipers, E.J., Nelis, G.F., Klinkenberg-Knol, E.C., Snel, P., Goldfain, D., Kolkman, J.J., Festen, H.P., Dent, J., Zeitoun, P., Havu, N., Lamm, M., and Walan, A. (2004) Cure of *Helicobacter pylori* infection in patients with reflux oesophagitis treated with long term omeprazole reverses gastritis without exacerbation of reflux disease: Results of a randomised controlled trial. *Gut* 53: 12-20.
- Kumagai, T., Malaty, H.M., Graham, D.Y., Hosogaya, S., Misawa, K., Furihata, K., Ota, H., Sei, C., Tanaka, E., Akamatsu, T., Shimizu, T., Kiyosawa, K., and Katsuyama, T. (1998) Acquisition versus loss of *Helicobacter pylori* infection in Japan: Results from an 8-year birth cohort study. *J Infect Dis* 178: 717-721.

- Lagergren, J. (2005) Adenocarcinoma of oesophagus: What exactly is the size of the problem and who is at risk? *Gut* 54 Suppl 1: i1-5.
- Lambert, J.R., Lin, S.K., Sievert, W., Nicholson, L., Schembri, M., and Guest, C. (1995) High prevalence of *Helicobacter pylori* antibodies in an institutionalized population: Evidence for person-to-person transmission. *Am J Gastroenterol* 90: 2167-2171.
- Laporte, R., Pernes, P., Pronnier, P., Gottrand, F., and Vincent, P. (2004) Acquisition of *Helicobacter pylori* infection after outbreaks of gastroenteritis: Prospective cohort survey in institutionalised young people. *BMJ* 329: 204-205.
- Leung, W.K., Siu, K.L., Kwok, C.K., Chan, S.Y., Sung, R., and Sung, J.J. (1999) Isolation of *Helicobacter pylori* from vomitus in children and its implication in gastro-oral transmission. *Am J Gastroenterol* 94: 2881-2884.
- Lin, L.F., Posfai, J., Roberts, R.J., and Kong, H. (2001) Comparative genomics of the restriction-modification systems in *Helicobacter pylori*. *Proc Natl Acad Sci U S A* 98: 2740-2745.
- Lin, S.K., Lambert, J.R., Schembri, M.A., Nicholson, L., and Korman, M.G. (1994) *Helicobacter pylori* prevalence in endoscopy and medical staff. J *Gastroenterol Hepatol* 9: 319-324.
- Lu, Y., Redlinger, T.E., Avitia, R., Galindo, A., and Goodman, K. (2002) Isolation and genotyping of *Helicobacter pylori* from untreated municipal wastewater. *Appl Environ Microbiol* 68: 1436-1439.
- Lundin, A., Björkholm, B., Kupershmidt, I., Unemo, M., Nilsson, P., Andersson, D.I., and Engstrand, L. (2005) Slow genetic divergence of *Helicobacter pylori* strains during long-term colonization. *Infect Immun* 73: 4818-4822.
- Luzza, F., Mancuso, M., Imeneo, M., Contaldo, A., Giancotti, L., Pensabene, L., Doldo, P., Liberto, M.C., Strisciuglio, P., Foca, A., Guandalini, S., and Pallone, F. (2000) Evidence favouring the gastro-oral route in the transmission of *Helicobacter pylori* infection in children. *Eur J Gastroenterol Hepatol* 12: 623-627.
- Magistà, A.M., Ierardi, E., Castellaneta, S., Miniello, V.L., Lionetti, E., Francavilla, A., Ros, P., Rigillo, N., Di Leo, A., and Francavilla, R. (2005) *Helicobacter pylori* status and symptom assessment two years after eradication in pediatric patients from a high prevalence area. J Pediatr Gastroenterol Nutr 40: 312-318.
- Magnusson, P.K.E., Enroth, H., Eriksson, I., Held, M., Nyrén, O., Engstrand, L., Hansson, L.E., and Gyllensten, U.B. (2001) Gastric cancer and human leukocyte antigen: Distinct DQ and DR alleles are associated with development of gastric cancer and infection by *Helicobacter pylori*. *Cancer Res* 61: 2684-2689.
- Mahdavi, J., Sondén, B., Hurtig, M., Olfat, F.O., Forsberg, L., Roche, N., Ångström, J., Larsson, T., Teneberg, S., Karlsson, K.A., Altraja, S., Wadström, T., Kersulyte, D., Berg, D.E., Dubois, A., Petersson, C., Magnusson, K.E., Norberg, T., Lindh, F., Lundskog, B.B., Arnqvist, A., Hammarström, L., and Borén, T. (2002) *Helicobacter pylori* SabA adhesin in persistent infection and chronic inflammation. *Science* 297: 573-578.

- Mahdavi, J., Boren, T., Vandenbroucke-Grauls, C., and Appelmelk, B.J. (2003) Limited role of lipopolysaccharide Lewis antigens in adherence of *Helicobacter pylori* to the human gastric epithelium. *Infect Immun* 71: 2876-2880.
- Malaty, H.M., Evans, D.G., Evans, D.J., Jr., and Graham, D.Y. (1992) *Helicobacter pylori* in Hispanics: Comparison with blacks and whites of similar age and socioeconomic class. *Gastroenterology* 103: 813-816.
- Malaty, H.M., Engstrand, L., Pedersen, N.L., and Graham, D.Y. (1994) *Helicobacter* pylori infection: Genetic and environmental influences. A study of twins. *Ann Intern Med* 120: 982-986.
- Malaty, H.M., Kim, J.G., Kim, S.D., and Graham, D.Y. (1996) Prevalence of *Helicobacter pylori* infection in Korean children: Inverse relation to socioeconomic status despite a uniformly high prevalence in adults. Am J Epidemiol 143: 257-262.
- Malaty, H.M., El-Kasabany, A., Graham, D.Y., Miller, C.C., Reddy, S.G., Srinivasan, S.R., Yamaoka, Y., and Berenson, G.S. (2002) Age at acquisition of *Helicobacter pylori* infection: A follow-up study from infancy to adulthood. *Lancet* 359: 931-935.
- Malfertheiner, P., Megraud, F., O'Morain, C., Hungin, A.P., Jones, R., Axon, A., Graham, D.Y., and Tytgat, G. (2002) Current concepts in the management of *Helicobacter pylori* infection: The Maastricht 2-2000 Consensus Report. *Aliment Pharmacol Ther* 16: 167-180.
- Malfertheiner, P., and Peitz, U. (2005) The interplay between *Helicobacter pylori*, gastro-oesophageal reflux disease, and intestinal metaplasia. *Gut* 54 Suppl 1: i13-20.
- Marchetti, M., and Rappuoli, R. (2002) Isogenic mutants of the *cag* pathogenicity island of *Helicobacter pylori* in the mouse model of infection: Effects on colonization efficiency. *Microbiology* 148: 1447-1456.
- Marshall, B.J., and Warren, J.R. (1984) Unidentified curved bacilli in the stomach of patients with gastritis and peptic ulceration. *Lancet* 1: 1311-1315.
- Marshall, B.J. (2001) *Helicobacter pylori* physiology and genetics. One hundred years of discovery and rediscovery of *Helicobacter pylori* and its association with peptic ulcer disease. Mobley, H.L.T., Mendz, G.L. and Hazell, S.L. (eds): ASM Press.
- Matsuda, R., Morizane, T., Tsunematsu, S., Kawana, I., and Tomiyama, M. (2002) *Helicobacter pylori* prevalence in dentists in Japan: A seroepidemiological study. *J Gastroenterol* 37: 255-259.
- McCallion, W.A., Murray, L.J., Bailie, A.G., Dalzell, A.M., O'Reilly, D.P., and Bamford, K.B. (1996) *Helicobacter pylori* infection in children: Relation with current household living conditions. *Gut* 39: 18-21.
- Mendall, M.A., Goggin, P.M., Molineaux, N., Levy, J., Toosy, T., Strachan, D., and Northfield, T.C. (1992) Childhood living conditions and *Helicobacter pylori* seropositivity in adult life. *Lancet* 339: 896-897.
- Merrell, D.S., Butler, S.M., Qadri, F., Dolganov, N.A., Alam, A., Cohen, M.B., Calderwood, S.B., Schoolnik, G.K., and Camilli, A. (2002) Host-induced epidemic spread of the cholera bacterium. *Nature* 417: 642-645.

- Miehlke, S., Genta, R.M., Graham, D.Y., and Go, M.F. (1999) Molecular relationships of *Helicobacter pylori* strains in a family with gastroduodenal disease. *Am J Gastroenterol* 94: 364-368.
- Milman, N., Rosenstock, S., Andersen, L., Jorgensen, T., and Bonnevie, O. (1998) Serum ferritin, hemoglobin, and *Helicobacter pylori* infection: A seroepidemiologic survey comprising 2794 Danish adults. *Gastroenterology* 115: 268-274.
- Mitchell, H.M., Li, Y.Y., Hu, P.J., Liu, Q., Chen, M., Du, G.G., Wang, Z.J., Lee, A., and Hazell, S.L. (1992) Epidemiology of *Helicobacter pylori* in southern China: Identification of early childhood as the critical period for acquisition. *J Infect Dis* 166: 149-153.
- Mitchell, H.M., Hu, P., Chi, Y., Chen, M.H., Li, Y.Y., and Hazell, S.L. (1998) A low rate of reinfection following effective therapy against *Helicobacter pylori* in a developing nation (China). *Gastroenterology* 114: 256-261.
- Moayyedi, P., Soo, S., Deeks, J., Delaney, B., Harris, A., Innes, M., Oakes, R., Wilson, S., Roalfe, A., Bennett, C., and Forman, D. (2005) Eradication of *Helicobacter pylori* for non-ulcer dyspepsia. *Cochrane Database Syst Rev*: CD002096.
- Mohammadi, M., Nedrud, J., Redline, R., Lycke, N., and Czinn, S.J. (1997) Murine CD4 T-cell response to Helicobacter infection: TH1 cells enhance gastritis and TH2 cells reduce bacterial load. *Gastroenterology* 113: 1848-1857.
- Molinari, M., Salio, M., Galli, C., Norais, N., Rappuoli, R., Lanzavecchia, A., and Montecucco, C. (1998) Selective inhibition of Ii-dependent antigen presentation by *Helicobacter pylori* toxin VacA. *J Exp Med* 187: 135-140.
- Nedenskov-Sörensen, P., Bukholm, G., and Bovre, K. (1990) Natural competence for genetic transformation in *Campylobacter pylori*. J Infect Dis 161: 365-366.
- NIH Consensus Conference (1994) *Helicobacter pylori* in peptic ulcer disease. NIH consensus development panel on *Helicobacter pylori* in peptic ulcer disease. *JAMA* 272: 65-69.
- Nilsson, C., Sillén, A., Eriksson, L., Strand, M.L., Enroth, H., Normark, S., Falk, P., and Engstrand, L. (2003) Correlation between *cag* pathogenicity island composition and *Helicobacter pylori*-associated gastroduodenal disease. *Infect Immun* 71: 6573-6581.
- Nomura, A.M., Perez-Perez, G.I., Lee, J., Stemmermann, G., and Blaser, M.J. (2002) Relation between *Helicobacter pylori cagA* status and risk of peptic ulcer disease. *Am J Epidemiol* 155: 1054-1059.
- Nurgalieva, Z.Z., Malaty, H.M., Graham, D.Y., Almuchambetova, R., Machmudova, A., Kapsultanova, D., Osato, M.S., Hollinger, F.B., and Zhangabylov, A. (2002) *Helicobacter pylori* infection in Kazakhstan: Effect of water source and household hygiene. *Am J Trop Med Hyg* 67: 201-206.
- Odenbreit, S., Puls, J., Sedlmaier, B., Gerland, E., Fischer, W., and Haas, R. (2000) Translocation of *Helicobacter pylori* CagA into gastric epithelial cells by type IV secretion. *Science* 287: 1497-1500.
- Park, C.Y., Cho, Y.K., Kodama, T., El-Zimaity, H.M., Osato, M.S., Graham, D.Y., and Yamaoka, Y. (2002) New serological assay for detection of putative *Helicobacter pylori* virulence factors. *J Clin Microbiol* 40: 4753-4756.

- Parsonnet, J. (1995) The incidence of *Helicobacter pylori* infection. *Aliment Pharmacol Ther* 9 Suppl 2: 45-51.
- Parsonnet, J., Friedman, G.D., Orentreich, N., and Vogelman, H. (1997) Risk for gastric cancer in people with CagA positive or CagA negative *Helicobacter pylori* infection. *Gut* 40: 297-301.
- Parsonnet, J., Shmuely, H., and Haggerty, T. (1999) Fecal and oral shedding of *Helicobacter pylori* from healthy infected adults. *JAMA* 282: 2240-2245.
- Passaro, D.J., Taylor, D.N., Meza, R., Cabrera, L., Gilman, R.H., and Parsonnet, J. (2001) Acute *Helicobacter pylori* infection is followed by an increase in diarrheal disease among Peruvian children. *Pediatrics* 108: E87.
- Perez-Perez, G.I., Witkin, S.S., Decker, M.D., and Blaser, M.J. (1991) Seroprevalence of *Helicobacter pylori* infection in couples. *J Clin Microbiol* 29: 642-644.
- Perez-Perez, G.I., Salomaa, A., Kosunen, T.U., Daverman, B., Rautelin, H., Aromaa, A., Knekt, P., and Blaser, M.J. (2002) Evidence that *cagA+ Helicobacter pylori* strains are disappearing more rapidly than *cagA-* strains. *Gut* 50: 295-298.
- Raymond, J., Thiberg, J.M., Chevalier, C., Kalach, N., Bergeret, M., Labigne, A., and Dauga, C. (2004) Genetic and transmission analysis of *Helicobacter pylori* strains within a family. *Emerg Infect Dis* 10: 1816-1821.
- Reilly, M., and Sullivan Pepe, M. (1995) A mean score method for missing and auxiliary covariate data in regression models. *Biometrika* 82: 299-314.
- Reilly, M. (1996) Optimal sampling strategies for two-stage studies. *Am J Epidemiol* 143: 92-100.
- Rocha, G.A., Rocha, A.M., Silva, L.D., Santos, A., Bocewicz, A.C., Queiroz Rd Rde, M., Bethony, J., Gazzinelli, A., Correa-Oliveira, R., and Queiroz, D.M. (2003) Transmission of *Helicobacter pylori* infection in families of preschool-aged children from Minas Gerais, Brazil. *Trop Med Int Health* 8: 987-991.
- Rothenbacher, D., Bode, G., Adler, G., and Brenner, H. (1998a) History of antibiotic treatment and prevalence of *H. pylori* infection among children: Results of a population-based study. *J Clin Epidemiol* 51: 267-271.
- Rothenbacher, D., Bode, G., Berg, G., Gommel, R., Gonser, T., Adler, G., and Brenner, H. (1998b) Prevalence and determinants of *Helicobacter pylori* infection in preschool children: A population-based study from Germany. *Int J Epidemiol* 27: 135-141.
- Rothenbacher, D., Blaser, M.J., Bode, G., and Brenner, H. (2000) Inverse relationship between gastric colonization of *Helicobacter pylori* and diarrheal illnesses in children: Results of a population-based cross-sectional study. *J Infect Dis* 182: 1446-1449.
- Rothenbacher, D., Bode, G., and Brenner, H. (2002a) History of breastfeeding and *Helicobacter pylori* infection in pre-school children: Results of a population-based study from Germany. *Int J Epidemiol* 31: 632-637.
- Rothenbacher, D., Winkler, M., Gonser, T., Adler, G., and Brenner, H. (2002b) Role of infected parents in transmission of *Helicobacter pylori* to their children. *Pediatr Infect Dis J* 21: 674-679.

- Rothenbacher, D., and Brenner, H. (2003) Burden of *Helicobacter pylori* and *H. pylori*-related diseases in developed countries: Recent developments and future implications. *Microbes Infect* 5: 693-703.
- Rothenbacher, D., Weyermann, M., Bode, G., Kulaksiz, M., Stahl, B., and Brenner,
 H. (2004) Role of Lewis A and Lewis B blood group antigens in *Helicobacter pylori* infection. *Helicobacter* 9: 324-329.
- Rowland, M., Kumar, D., Daly, L., O'Connor, P., Vaughan, D., and Drumm, B. (1999) Low rates of *Helicobacter pylori* reinfection in children. *Gastroenterology* 117: 336-341.
- Ruggiero, P., Peppoloni, S., Rappuoli, R., and Del Giudice, G. (2003) The quest for a vaccine against *Helicobacter pylori*: How to move from mouse to man? *Microbes Infect* 5: 749-756.
- Rupnow, M.F., Shachter, R.D., Owens, D.K., and Parsonnet, J. (2000) A dynamic transmission model for predicting trends in *Helicobacter pylori* and associated diseases in the United States. *Emerg Infect Dis* 6: 228-237.
- Salama, N., Guillemin, K., McDaniel, T.K., Sherlock, G., Tompkins, L., and Falkow, S. (2000) A whole-genome microarray reveals genetic diversity among *Helicobacter pylori* strains. *Proc Natl Acad Sci U S A* 97: 14668-14673.
- Salama, N.R., Shepherd, B., and Falkow, S. (2004) Global transposon mutagenesis and essential gene analysis of *Helicobacter pylori*. J Bacteriol 186: 7926-7935.
- Salaün, L., Ayraud, S., and Saunders, N.J. (2005) Phase variation mediated niche adaptation during prolonged experimental murine infection with *Helicobacter pylori*. *Microbiology* 151: 917-923.
- Salgueiro, J., Zubillaga, M., Goldman, C., Barrado, A., Martinez Sarrasague, M., Leonardi, N., and Boccio, J. (2004) Review article: Is there a link between micronutrient malnutrition and *Helicobacter pylori* infection? *Aliment Pharmacol Ther* 20: 1029-1034.
- Segal, E.D., Cha, J., Lo, J., Falkow, S., and Tompkins, L.S. (1999) Altered states: Involvement of phosphorylated CagA in the induction of host cellular growth changes by *Helicobacter pylori*. *Proc Natl Acad Sci U S A* 96: 14559-14564.
- Silicon Genetics (2004) GeneSpring user manual version 7.0. Agilent Technologies/Silicon Genetics. USA.
- Simán, J.H., Engstrand, L., Berglund, G., Florén, C.H., and Forsgren, A. (2005) Evaluation of western blot CagA seropositivity in *Helicobacter pylori*seropositive and -seronegative subjects. *Clin Diagn Lab Immunol* 12: 304-309.
- Simoons-Smit, I.M., Appelmelk, B.J., Verboom, T., Negrini, R., Penner, J.L., Aspinall, G.O., Moran, A.P., Fei, S.F., Shi, B.S., Rudnica, W., Savio, A., and de Graaff, J. (1996) Typing of *Helicobacter pylori* with monoclonal antibodies against Lewis antigens in lipopolysaccharide. *J Clin Microbiol* 34: 2196-2200.
- Sjölund, M., Wreiber, K., Andersson, D.I., Blaser, M.J., and Engstrand, L. (2003) Long-term persistence of resistant *Enterococcus* species after antibiotics to eradicate *Helicobacter pylori*. *Ann Intern Med* 139: 483-487.

- Solnick, J.V., Hansen, L.M., Salama, N.R., Boonjakuakul, J.K., and Syvanen, M. (2004) Modification of *Helicobacter pylori* outer membrane protein expression during experimental infection of rhesus macaques. *Proc Natl Acad Sci U S A* 101: 2106-2111.
- Soto, G., Bautista, C.T., Roth, D.E., Gilman, R.H., Velapatino, B., Ogura, M., Dailide, G., Razuri, M., Meza, R., Katz, U., Monath, T.P., Berg, D.E., and Taylor, D.N. (2003) *Helicobacter pylori* reinfection is common in Peruvian adults after antibiotic eradication therapy. *J Infect Dis* 188: 1263-1275.
- Statistics Sweden (1995) Swedish socioeconomic classification. Reports on statistical co-ordination 1982:4 (Swedish). Statistics Sweden. Sweden.
- Suerbaum, S., Smith, J.M., Bapumia, K., Morelli, G., Smith, N.H., Kunstmann, E., Dyrek, I., and Achtman, M. (1998) Free recombination within *Helicobacter pylori*. *Proc Natl Acad Sci U S A* 95: 12619-12624.
- Suerbaum, S., and Michetti, P. (2002) *Helicobacter pylori* infection. *N Engl J Med* 347: 1175-1186.
- Suto, H., Zhang, M., and Berg, D.E. (2005) Age-dependent changes in susceptibility of suckling mice to individual strains of *Helicobacter pylori*. *Infect Immun* 73: 1232-1234.
- Sörberg, M., Engstrand, L., Ström, M., Jönsson, K.Å., Jörbeck, H., and Granström, M. (1997) The diagnostic value of enzyme immunoassay and immunoblot in monitoring eradication of *Helicobacter pylori*. Scand J Infect Dis 29: 147-151.
- Taylor, D.E., Rasko, D.A., Sherburne, R., Ho, C., and Jewell, L.D. (1998) Lack of correlation between Lewis antigen expression by *Helicobacter pylori* and gastric epithelial cells in infected patients. *Gastroenterology* 115: 1113-1122.
- Taylor, N.S., Fox, J.G., Akopyants, N.S., Berg, D.E., Thompson, N., Shames, B., Yan, L., Fontham, E., Janney, F., Hunter, F.M., and et al. (1995) Long-term colonization with single and multiple strains of *Helicobacter pylori* assessed by DNA fingerprinting. *J Clin Microbiol* 33: 918-923.
- Terry, K., Williams, S.M., Connolly, L., and Ottemann, K.M. (2005) Chemotaxis plays multiple roles during *Helicobacter pylori* animal infection. *Infect Immun* 73: 803-811.
- Testerman, T.L., McGee, D.J., and Mobley, H.L.T. (2001) *Helicobacter pylori* physiology and genetics. Adherence and colonization. Mobley, H.L.T., Mendz, G.L. and Hazell, S.L. (eds): ASM Press. USA.
- Thomas, J.E., Gibson, G.R., Darboe, M.K., Dale, A., and Weaver, L.T. (1992) Isolation of *Helicobacter pylori* from human faeces. *Lancet* 340: 1194-1195.
- Thomas, J.E., Austin, S., Dale, A., McClean, P., Harding, M., Coward, W.A., and Weaver, L.T. (1993) Protection by human milk IgA against *Helicobacter pylori* infection in infancy. *Lancet* 342: 121.
- Tindberg, Y., Bengtsson, C., Bergström, M., and Granström, M. (2001a) The accuracy of serologic diagnosis of *Helicobacter pylori* infection in schoolaged children of mixed ethnicity. *Helicobacter* 6: 24-30.
- Tindberg, Y., Bengtsson, C., Granath, F., Blennow, M., Nyrén, O., and Granström, M. (2001b) *Helicobacter pylori* infection in Swedish school children: Lack of

evidence of child-to-child transmission outside the family. *Gastroenterology* 121: 310-316.

- Tomasini, M.L., Zanussi, S., Sozzi, M., Tedeschi, R., Basaglia, G., and De Paoli, P. (2003) Heterogeneity of *cag* genotypes in *Helicobacter pylori* isolates from human biopsy specimens. *J Clin Microbiol* 41: 976-980.
- Tomb, J.F., White, O., Kerlavage, A.R., Clayton, R.A., Sutton, G.G., Fleischmann, R.D., Ketchum, K.A., Klenk, H.P., Gill, S., Dougherty, B.A., Nelson, K., Quackenbush, J., Zhou, L., Kirkness, E.F., Peterson, S., Loftus, B., Richardson, D., Dodson, R., Khalak, H.G., Glodek, A., McKenney, K., Fitzegerald, L.M., Lee, N., Adams, M.D., and Venter, J.C. (1997) The complete genome sequence of the gastric pathogen *Helicobacter pylori*. *Nature* 388: 539-547.
- Torres, J., Perez-Perez, G., Goodman, K.J., Atherton, J.C., Gold, B.D., Harris, P.R., la Garza, A.M., Guarner, J., and Munoz, O. (2000) A comprehensive review of the natural history of *Helicobacter pylori* infection in children. *Arch Med Res* 31: 431-469.
- Tsai, C.J., Perry, S., Sanchez, L., and Parsonnet, J. (2005) *Helicobacter pylori* infection in different generations of hispanics in the San Francisco bay area. *Am J Epidemiol* 162: 351-357.
- Uemura, N., Okamoto, S., Yamamoto, S., Matsumura, N., Yamaguchi, S., Yamakido, M., Taniyama, K., Sasaki, N., and Schlemper, R.J. (2001) *Helicobacter pylori* infection and the development of gastric cancer. *N Engl J Med* 345: 784-789.
- Umlauft, F., Keeffe, E.B., Offner, F., Weiss, G., Feichtinger, H., Lehmann, E., Kilga-Nogler, S., Schwab, G., Propst, A., Grussnewald, K., and Judmaier, G. (1996) *Helicobacter pylori* infection and blood group antigens: Lack of clinical association. *Am J Gastroenterol* 91: 2135-2138.
- van Belkum, A., Struelens, M., de Visser, A., Verbrugh, H., and Tibayrenc, M. (2001) Role of genomic typing in taxonomy, evolutionary genetics, and microbial epidemiology. *Clin Microbiol Rev* 14: 547-560.
- van der Ende, A., Rauws, E.A., Feller, M., Mulder, C.J., Tytgat, G.N., and Dankert, J. (1996) Heterogeneous *Helicobacter pylori* isolates from members of a family with a history of peptic ulcer disease. *Gastroenterology* 111: 638-647.
- Wang, J., Brooks, E.G., Bamford, K.B., Denning, T.L., Pappo, J., and Ernst, P.B. (2001) Negative selection of T cells by *Helicobacter pylori* as a model for bacterial strain selection by immune evasion. *J Immunol* 167: 926-934.
- Webb, P.M., Knight, T., Greaves, S., Wilson, A., Newell, D.G., Elder, J., and Forman, D. (1994) Relation between infection with *Helicobacter pylori* and living conditions in childhood: Evidence for person to person transmission in early life. *BMJ* 308: 750-753.
- Veldhuyzen van Zanten, S.J., Pollak, P.T., Best, L.M., Bezanson, G.S., and Marrie, T. (1994) Increasing prevalence of *Helicobacter pylori* infection with age: Continuous risk of infection in adults rather than cohort effect. *J Infect Dis* 169: 434-437.
- Wheeldon, T.U., Hoang, T.T., Phung, D.C., Björkman, A., Granström, M., and Sörberg, M. (2005) Long-term follow-up of *Helicobacter pylori* eradication

therapy in Vietnam: Reinfection and clinical outcome. *Aliment Pharmacol Ther* 21: 1047-1053.

- Viala, J., Chaput, C., Boneca, I.G., Cardona, A., Girardin, S.E., Moran, A.P., Athman, R., Memet, S., Huerre, M.R., Coyle, A.J., DiStefano, P.S., Sansonetti, P.J., Labigne, A., Bertin, J., Philpott, D.J., and Ferrero, R.L. (2004) Nod1 responds to peptidoglycan delivered by the *Helicobacter pylori cag* pathogenicity island. *Nat Immunol* 5: 1166-1174.
- Wirth, H.P., Yang, M., Peek, R.M., Jr., Tham, K.T., and Blaser, M.J. (1997) *Helicobacter pylori* Lewis expression is related to the host Lewis phenotype. *Gastroenterology* 113: 1091-1098.
- Wirth, H.P., Yang, M., Peek, R.M., Jr., Hook-Nikanne, J., Fried, M., and Blaser, M.J. (1999) Phenotypic diversity in Lewis expression of *Helicobacter pylori* isolates from the same host. *J Lab Clin Med* 133: 488-500.
- Wirth, T., Wang, X., Linz, B., Novick, R.P., Lum, J.K., Blaser, M., Morelli, G., Falush, D., and Achtman, M. (2004) Distinguishing human ethnic groups by means of sequences from *Helicobacter pylori*: Lessons from Ladakh. *Proc Natl Acad Sci U S A* 101: 4746-4751.
- Wong, B.C., Lam, S.K., Wong, W.M., Chen, J.S., Zheng, T.T., Feng, R.E., Lai, K.C., Hu, W.H., Yuen, S.T., Leung, S.Y., Fong, D.Y., Ho, J., and Ching, C.K. (2004) *Helicobacter pylori* eradication to prevent gastric cancer in a high-risk region of China: A randomized controlled trial. *JAMA* 291: 187-194.
- Woodward, M., Morrison, C., and McColl, K. (2000) An investigation into factors associated with *Helicobacter pylori* infection. *J Clin Epidemiol* 53: 175-181.
- Xiang, Z., Censini, S., Bayeli, P.F., Telford, J.L., Figura, N., Rappuoli, R., and Covacci, A. (1995) Analysis of expression of CagA and VacA virulence factors in 43 strains of *Helicobacter pylori* reveals that clinical isolates can be divided into two major types and that CagA is not necessary for expression of the vacuolating cytotoxin. *Infect Immun* 63: 94-98.
- Xu, Q., Morgan, R.D., Roberts, R.J., and Blaser, M.J. (2000) Identification of type II restriction and modification systems in *Helicobacter pylori* reveals their substantial diversity among strains. *Proc Natl Acad Sci U S A* 97: 9671-9676.
- Yamaoka, Y., Kita, M., Kodama, T., Sawai, N., and Imanishi, J. (1996) *Helicobacter* pylori cagA gene and expression of cytokine messenger RNA in gastric mucosa. *Gastroenterology* 110: 1744-1752.
- Yamaoka, Y., Kodama, T., Graham, D.Y., and Kashima, K. (1998) Comparison of four serological tests to determine the CagA or VacA status of *Helicobacter pylori* strains. J Clin Microbiol 36: 3433-3434.
- Yamaoka, Y., Kita, M., Kodama, T., Imamura, S., Ohno, T., Sawai, N., Ishimaru, A., Imanishi, J., and Graham, D.Y. (2002) *Helicobacter pylori* infection in mice: Role of outer membrane proteins in colonization and inflammation. *Gastroenterology* 123: 1992-2004.
- Ye, W., Held, M., Enroth, H., Kraaz, W., Engstrand, L., and Nyrén, O. (2005) Histology and culture results among subjects with antibodies to CagA but no evidence of *Helicobacter pylori* infection with IgG ELISA. *Scand J Gastroenterol* 40: 312-318.

Zambon, C.F., Navaglia, F., Basso, D., Rugge, M., and Plebani, M. (2003) *Helicobacter pylori babA2, cagA*, and s1 *vacA* genes work synergistically in causing intestinal metaplasia. *J Clin Pathol* 56: 287-291.